

FORM PTO-1390 (Modified)  
(REV 10-95)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

## TRANSMITTAL LETTER TO THE UNITED STATES

01123.0004

DESIGNATED/ELECTED OFFICE (DO/EO/US)

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

CONCERNING A FILING UNDER 35 U.S.C. 371

09/509712

INTERNATIONAL APPLICATION NO.  
PCT/US98/21276INTERNATIONAL FILING DATE  
8 October 1998PRIORITY DATE CLAIMED  
10 October 1997

## TITLE OF INVENTION

MAMMALIAN GENES INVOLVED IN VIRAL INFECTION AND TUMOR SUPPRESSION

## APPLICANT(S) FOR DO/EO/US

RUBIN, Donald H.; ORGAN, Edward L.; DUBOIS, Raymond N.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
  - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☐ has been transmitted by the International Bureau.
  - c. ☒ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☐ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
  - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☐ have been transmitted by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
10. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☐ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

## Items 13 to 18 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.  
A **SECOND** or **SUBSEQUENT** preliminary amendment.
16. ☐ A substitute specification.
17. ☐ A change of power of attorney and/or address letter.
18. ☒ Certificate of Mailing by Express Mail
19. ☒ Other items or information:

Check in the amount of \$943.00 for filing fees; return-receipt postcard; Certificate of Express Mailing No. EL348124625US; Verified Statements Claiming Small Entity Status signed by Vanderbilt University and by Avatar BioSci, Inc.

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR <div style="font-size: 2em; font-weight: bold; text-align: center;">09/509712</div>		INTERNATIONAL APPLICATION NO. PCT/US98/21276		ATTORNEY'S DOCKET NUMBER 01123.0004	
20. The following fees are submitted: <b>BASIC NATIONAL FEE ( 37 CFR 1.492 (a) (1) - (5)) :</b> <div style="margin-left: 20px;"><input checked="" type="checkbox"/> Search Report has been prepared by the EPO or JPO ..... <span style="float: right;">\$840.00</span> <input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) ..... <span style="float: right;">\$670.00</span> <input type="checkbox"/> No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)) ..... <span style="float: right;">\$760.00</span> <input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2) paid to USPTO ..... <span style="float: right;">\$970.00</span> <input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) ..... <span style="float: right;">\$96.00</span></div> <div style="text-align: center; margin-top: 10px;"><b>ENTER APPROPRIATE BASIC FEE AMOUNT =</b></div>				<b>CALCULATIONS PTO USE ONLY</b>	
Surcharge of <b>\$130.00</b> for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).				<div style="border: 1px solid black; padding: 2px;">\$840.00</div> <div style="border: 1px solid black; padding: 2px;">\$0.00</div>	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	29 - 20 =	9	x \$18.00	<b>\$162.00</b>	
Independent claims	11 - 3 =	8	x \$78.00	<b>\$624.00</b>	
Multiple Dependent Claims (check if applicable).			<input checked="" type="checkbox"/>	<b>\$260.00</b>	
<b>TOTAL OF ABOVE CALCULATIONS =</b>				<b>\$1,886.00</b>	
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable).			<input checked="" type="checkbox"/>	<b>\$943.00</b>	
<b>SUBTOTAL =</b>				<b>\$943.00</b>	
Processing fee of <b>\$130.00</b> for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).				<b>\$0.00</b>	
<b>TOTAL NATIONAL FEE =</b>				<b>\$943.00</b>	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).			<input type="checkbox"/>	<b>\$0.00</b>	
<b>TOTAL FEES ENCLOSED =</b>				<b>\$943.00</b>	
				Amount to be: refunded	\$
				charged	\$
<div style="margin-bottom: 10px;"><input checked="" type="checkbox"/> A check in the amount of <b>\$943.00</b> to cover the above fees is enclosed.</div> <div style="margin-bottom: 10px;"><input type="checkbox"/> Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees. A duplicate copy of this sheet is enclosed.</div> <div><input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. <b>14-0629</b> A duplicate copy of this sheet is enclosed.</div>					
<p><b>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.</b></p> <p><b>SEND ALL CORRESPONDENCE TO:</b></p> <div style="display: flex; justify-content: space-between;"><div style="width: 45%; border: 1px solid black; padding: 5px;"><p>Janice A. Kimpel, Ph.D. NEEDLE &amp; ROSENBERG, P.C. 127 Peachtree Street, N.E., Suite 1200 Atlanta, Georgia 30303 U.S.A. (404) 688 0770</p></div><div style="width: 50%; text-align: right;"><div style="margin-bottom: 10px;"> SIGNATURE</div><div style="margin-bottom: 10px;">Janice A. Kimpel NAME</div><div style="margin-bottom: 10px;">42,734 REGISTRATION NUMBER</div><div>March 30, 2000 DATE</div></div></div>					

CERTIFICATE OF EXPRESS MAILING

I hereby certify that the filing of the U.S. National Phase in the names of Rubin *et al.*, consisting of: Transmittal Letter to the United States Designated/Elected Office (DO/EO/US) Concerning a Filing Under 35 U.S.C. 371 (2 pages), a check in the amount of \$943.00 for filing fee, Verified Statement Claiming Small Entity Status by Vanderbilt University (2 pages), Verified Statement Claiming Small Entity Status by Avatar BioSci, Inc. (2 pages), Declaration/Power of Attorney by Rubin, Organ & Dubois and return-receipt postcard are being deposited with the United States Postal Service as Express Mail No. EL348124625US in an envelope addressed to: Box PCT, Assistant Commissioner for Patents, Washington, D.C., 20231, on this **30th** day of **March 2000**.

  
\_\_\_\_\_  
Everardo McFarlane

3-30-2000  
Date

ATTORNEY DOCKET NO. 01123.0004

PAGE 1 OF 2

Applicant or Patentee: Donald H. Rubin, Edward L. Organ and Raymond N. Dubois

For: "MAMMALIAN GENES INVOLVED IN VIRAL  
INFECTION AND TUMOR SUPPRESSION"

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY  
STATUS (37 CFR 1.9(f) and 1.27(d)) - SMALL BUSINESS CONCERN**

I hereby declare that I am

- ☐ the owner of the small business concern identified below:  
☒ an official of the small business concern empowered to act on behalf of the  
concern identified below:

NAME OF CONCERN: Avatar Biosci, Inc.  
ADDRESS OF CONCERN: 1937 Edenbridge Way  
Nashville, TN 37215

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18(d), for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention, entitled "MAMMALIAN GENES INVOLVED IN VIRAL INFECTION AND TUMOR SUPPRESSION" by inventor(s) Donald H. Rubin, Edward L. Organ and Raymond N. Dubois described in the specification filed concurrently herewith.

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below\* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9(d) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).



\*NOTE: Separate verified statements are required for each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME: VANDERBILT UNIVERSITY  
ADDRESS: Office of Technology Transfer  
1207 17<sup>th</sup> Ave. S., Suite 210  
Nashville, TN 37212

☐ INDIVIDUAL ☒ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING:

Donald H Rubin MD

TITLE IN ORGANIZATION:

President

ADDRESS OF PERSON SIGNING:

Avatar Biosci, Inc.  
1937 Edenbridge Way  
Nashville, TN 37215

SIGNATURE

Donald H Rubin

DATE 3.28.00

ATTORNEY DOCKET NO. 01123.0004

**APPLICANT OR PATENTEE:** Donald H. Rubin, Edward L. Organ and Raymond N. Dubois  
**FOR:** "MAMMALIAN GENES INVOLVED IN VIRAL  
 INFECTION AND TUMOR SUPPRESSION"

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY  
 STATUS (37 CFR 1.9(f) and 1.27(d)) - NONPROFIT ORGANIZATION**

I hereby declare that I am an official empowered to act on behalf of the nonprofit organization identified below:

**NAME OF ORGANIZATION:** VANDERBILT UNIVERSITY  
**ADDRESS OF ORGANIZATION:** Office of Technology Transfer  
 1207 17<sup>th</sup> Ave. S., Suite 210  
 Nashville, TN 37212

**TYPE OF ORGANIZATION:**

- ☒ University or other institution of higher education  
 (Name of state: Tennessee  
 (Citation of statute: \_\_\_\_\_ )
- ☐ Tax exempt under Internal Revenue Service Code (26 USC 501(a) and 501(c)(3))
- ☐ Non-profit scientific or educational under statute of state of the United States of America  
 (Name of state \_\_\_\_\_ )  
 (Citation of statute \_\_\_\_\_ )
- ☐ Would qualify as tax exempt under Internal Revenue Service Code (26 USC 501(a) and 501(c)(3)) if it were located in The United States of America
- ☐ Would qualify as nonprofit scientific or educational under statute of state of The United States of America if it were located in The United States of America  
 (Name of state \_\_\_\_\_ )  
 (Citation of statute \_\_\_\_\_ )

I hereby declare that the nonprofit organization identified above qualifies as a nonprofit organization as defined in 37 CFR 1.9 (e) for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code with regard to the invention entitled "MAMMALIAN GENES INVOLVED IN VIRAL INFECTION AND TUMOR SUPPRESSION" by inventor(s) Donald H. Rubin, Edward L. Organ and Raymond N. Dubois filed concurrently herewith.

I hereby declare that rights under contract or law have been conveyed to and remain with the nonprofit organization with regard to the above identified invention.

If the rights held by the nonprofit organization are not exclusive, each individual, concern or organization having rights to the invention is listed below\* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR

1.9(d) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

\*NOTE: Separate verified statements are required for each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME Avatar Biosci, Inc.

ADDRESS 1937 Edenbridge Way  
Nashville, TN 37215

☐ INDIVIDUAL ☒ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING:

JANIS ELSNER

TITLE IN ORGANIZATION:

ASSOC. DIRECTOR

ADDRESS OF PERSON SIGNING:

Vanderbilt University  
Office of Technology Transfer  
1207 17<sup>th</sup> Ave. S., Suite 210  
Nashville, TN 37212

SIGNATURE:

Janis Elsner

DATE: Mar 29, 2000

430 Rec'd PCT/PTO 3-1 MAR 2000

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

For: **“MAMMALIAN GENES INVOLVED  
IN VIRAL INFECTION AND  
TUMOR SUPPRESSION”**

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Examiner: Unassigned

**PRELIMINARY AMENDMENT**

NEEDLE & ROSENBERG, P.C.  
Suite 1200, The Candler Building  
127 Peachtree Street, N.E.  
Atlanta, Georgia 30303-1811

March 30, 2000

Concurrently with the filing of this application, Applicants respectfully request entry of the following amendment:

**IN THE SPECIFICATION:**

On Page 1, at line 3 immediately after the title “Mammalian Genes Involved in Viral Infection and Tumor Suppression”, please insert the following:

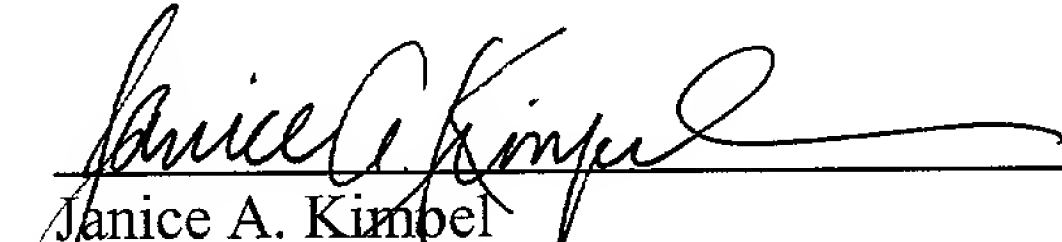
“This invention was made with partial government support under National Institutes of Health Grant No. CA68283 and a grant from the Department of Veterans Affairs. The United States Government has certain rights in the invention.”

ATTORNEY DOCKET NO. 01123.0004  
PATENT

No fee is believed due; however, the Commissioner is hereby authorized to charge any deficiency to Deposit Account No. 14-0629.

Respectfully submitted,

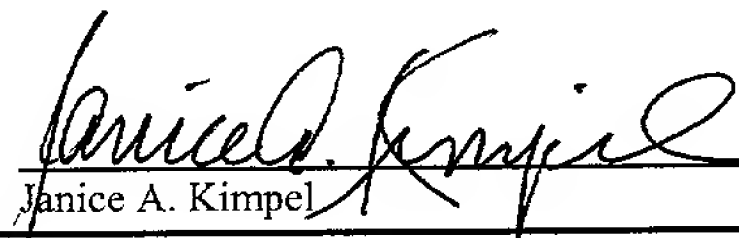
NEEDLE & ROSENBERG, P.C.

  
\_\_\_\_\_  
Janice A. Kimpel  
Registration No. 42,734

NEEDLE & ROSENBERG, P.C.  
Suite 1200, The Candler Building  
127 Peachtree Street, N.E.  
Atlanta, Georgia 30303-1811  
404/688-0770

CERTIFICATE OF EXPRESS MAILING

I hereby certify that this correspondence is being deposited with the United States Postal Service as Express Mailing No. EL348124625US in an envelope addressed to: Assistant Commissioner for Patents, Box PCT, Washington, D.C. 20231, on this 30th day of March, 2000.

  
\_\_\_\_\_  
Janice A. Kimpel

March 30, 2000  
\_\_\_\_\_  
Date

MAMMALIAN GENES INVOLVED IN VIRAL INFECTION  
AND TUMOR SUPPRESSION

**BACKGROUND**

**Field of the Invention**

The present invention provides methods of identifying cellular genes used for  
5 viral growth or for tumor progression. Thus, the present invention relates to nucleic  
acids related to and methods of reducing or preventing viral infection and for suppressing  
tumor progression. The invention also relates to methods for screening for additional  
such genes.

**Background art**

10 Various projects have been directed toward isolating and sequencing the genome  
of various animals, notably the human. However, most methodologies provide  
nucleotide sequences for which no function is linked or even suggested, thus limiting the  
immediate usefulness of such data.

The present invention, in contrast, provides methods of screening only for nucleic  
15 acids that are involved in a specific process, *i.e.*, viral infection or tumor progression.  
For viral infection, the nucleic acids isolated are useful in treatments for these processes  
because by this method only nucleic acids which are also nonessential to the cell are  
isolated. Such methods are highly useful, since they ascribe a function to each isolated  
gene, and thus the isolated nucleic acids can immediately be utilized in various specific  
20 methods and procedures.

For, example, the present invention provides methods of isolating nucleic acids  
encoding gene products used for viral infection, but nonessential to the cell. Viral  
infections are significant causes of human morbidity and mortality. Understanding the  
molecular mechanisms of such infections will lead to new approaches in their treatment  
25 and control.

Viruses can establish a variety of types of infection. These infections can be  
generally classified as lytic or persistent, though some lytic infections are considered  
persistent. Generally, persistent infections fall into two categories: (1) chronic  
(productive) infection, *i.e.*, infection wherein infectious virus is present and can be  
30 recovered by traditional biological methods and (2) latent infection, *i.e.*, infection



wherein viral genome is present in the cell but infectious virus is generally not produced except during intermittent episodes of reactivation. Persistence generally involves stages of both productive and latent infection.

Lytic infections can also persist under conditions where only a small fraction of the total cells are infected (smoldering (cycling) infection). The few infected cells release virus and are killed, but the progeny virus again only infect a small number of the total cells. Examples of such smoldering infections include the persistence of lactic dehydrogenase virus in mice (Mahy, B.W.J., *Br. Med. Bull.* 41: 50-55 (1985)) and adenovirus infection in humans (Porter, D.D. pp. 784-790 in Baron, S., ed. *Medical Microbiology* 2d ed. (Addison-Wesley, Menlo Park, CA 1985)).

Furthermore, a virus may be lytic for some cell types but not for others. For example, evidence suggests that human immunodeficiency virus (HIV) is more lytic for T cells than for monocytes/macrophages, and therefore can result in a productive infection of T cells that can result in cell death, whereas HIV-infected mononuclear phagocytes may produce virus for considerable periods of time without cell lysis. (Klatzmann, et al. *Science* 225:59-62 (1984); Koyanagi, et al. *Science* 241:1673-1675 (1988); Sattentau, et al. *Cell* 52:631-633 (1988)).

Traditional treatments for viral infection include pharmaceuticals aimed at specific virus derived proteins, such as HIV protease or reverse transcriptase, or recombinant (cloned) immune modulators (host derived), such as the interferons. However, the current methods have several limitations and drawbacks which include high rates of viral mutations which render anti-viral pharmaceuticals ineffective. For immune modulators, limited effectiveness, limiting side effects, a lack of specificity all limit the general applicability of these agents. Also the rate of success with current antivirals and immune-modulators has been disappointing.

One aspect of the current invention focuses on isolating genes that are not essential for cellular survival when disrupted in one or both alleles, but which are required for virus replication. This may occur with a dose effect, in which one allele knock-out may confer the phenotype of virus resistance for the cell. As targets for therapeutic intervention, inhibition of these cellular gene products, including: proteins, parts of proteins (modification enzymes that include, but are not restricted to glycosylation, lipid modifiers [myriolate, etc.]), lipids, transcription elements and RNA

regulatory molecules, may be less likely to have profound toxic side effects and virus mutation is less likely to overcome the 'block' to replicate successfully.

The present invention provides a significant improvement over previous methods of attempted therapeutic intervention against viral infection by addressing the cellular genes required by the virus for growth. Therefore, the present invention also provides an innovative therapeutic approach to intervention in viral infection by providing methods to treat viruses by inhibiting the cellular genes necessary for viral infection. Because these genes, by virtue of the means by which they are originally detected, are nonessential to the cell's survival at a level of expression necessary to inhibit virus replication, these treatment methods can be used in a subject without serious detrimental effects to the subject, as has been found with previous methods. The present invention also provides the surprising discovery that virally infected cells are dependent upon a factor in serum to survive. Therefore, the present invention also provides a method for treating viral infection by inhibiting this serum survival factor. Finally, these discoveries also provide a novel method for removing virally infected cells from a cell culture by removing, inhibiting or disrupting this serum survival factor in the culture so that non-infected cells selectively survive.

The selection of tumor suppressor gene(s) has become an important area in the discovery of new target for therapeutic intervention of cancer. Since the discovery that cells are restricted from promiscuous entry into the cell cycle by specific genes that are capable of suppressing a 'transformed' phenotype, considerable time has been invested in the discovery of such genes. Some of these genes include the gene associated by rhabdomyosarcoma (Rb) and the p53 (apoptosis related) encoding gene. The present invention provides a method, using gene-trapping, to select cell lines that have a transformed phenotype from cells that are not transformed and to isolate from these cells a gene that can suppress a malignant, or transformed, phenotype. Thus, by the nature of the isolation process, a function is associated with the isolated genes. The capacity to select quickly tumor suppressor genes can provide unique targets in the process of treating or preventing, and even for diagnostic testing of, cancer.

## DETAILED DESCRIPTION OF THE INVENTION

The present invention utilizes a "gene trap" method along with a selection process to identify and isolate nucleic acids from genes associated with a particular function. Specifically, it provides a means of isolating cellular genes necessary for viral infection but not essential for the cell's survival, and it provides a means of isolating cellular genes that suppress tumor progression.

The present invention also provides a core discovery that virally infected cells become dependent upon at least one factor present in serum for survival, whereas non-infected cells do not exhibit this dependence. This core discovery has been utilized in the present invention in several ways. First, inhibition of the "serum survival factor" can be utilized to eradicate persistently virally infected cells from populations of non-infected cells. Inhibition of this factor can also be used to treat virus infection in a subject, as further described herein. Additionally, inhibition of or withdrawal of the serum survival factor in tissue culture allows for the detection of cellular genes required for viral replication yet nonessential for an uninfected cell to survive. The present invention further provides several such cellular genes, as well as methods of treating viral infections by inhibiting the functioning of such genes.

The invention also provides cellular genes whose overexpression is associated with inhibition of viral growth and/or reproduction.

The present method provides several cellular genes that are necessary for viral growth in the cell but are not essential for the cell to survive. These genes are important for lytic and persistent infection by viruses. These genes were isolated by generating gene trap libraries by infecting cells with a retrovirus gene trap vector, selecting for cells in which a gene trap event occurred (*i.e.*, in which the vector had inserted such that the promoterless marker gene was inserted such that a cellular promoter promotes transcription of the marker gene, *i.e.*, inserted into a functioning gene), starving the cells of serum, infecting the selected cells with the virus of choice while continuing serum starvation, and adding back serum to allow visible colonies to develop, which colonies were cloned by limiting dilution. Genes into which the retrovirus gene trap vector inserted were then isolated from the colonies using probes specific for the retrovirus gene trap vector. Thus nucleic acids isolated by this method are isolated portions of genes. Additionally, utilizing this method, several cellular genes were isolated whose

overexpression prevents viral infection or tumor growth, and they provide methods of treating viral infection or tumor growth/suppression by overexpression of these genes.

Thus the present invention provides a method of identifying a cellular gene necessary for viral growth in a cell and nonessential for cellular survival, comprising (a) transferring into a cell culture, *e.g.* growing in serum-containing medium, a vector encoding a selective marker gene lacking a functional promoter, (b) selecting cells expressing the marker gene, (c) removing serum from the culture medium, (d) infecting the cell culture with the virus, and (e) isolating from the surviving cells a cellular gene within which the marker gene is inserted, thereby identifying a gene necessary for viral growth in a cell and nonessential for cellular survival. The present invention also provides a method of identifying a cellular gene used for viral growth in a cell and nonessential for cellular survival, comprising (a) transferring into a cell culture growing in serum-containing medium a vector encoding a selective marker gene lacking a functional promoter, (b) selecting cells expressing the marker gene, (c) removing serum from the culture medium, (d) infecting the cell culture with the virus, and (e) isolating from the surviving cells a cellular gene within which the marker gene is inserted, thereby identifying a gene necessary for viral growth in a cell and nonessential for cellular survival or a gene whose overexpression prevents viral reproduction but is not fatal to the survival to the cell. In any selected cell type, such as Chinese hamster ovary cells, one can readily determine if serum starvation is required for selection. If it is not, serum starvation may be eliminated from the steps.

Alternatively, instead of removing serum from the culture medium, a serum factor required by the virus for growth can be inhibited, such as by the administration of an antibody that specifically binds that factor. Furthermore, if it is believed that there are no persistently infected cells in the culture, the serum starvation step can be eliminated and the cells grown in usual medium for the cell type. If serum starvation is used, it can be continued for a time after the culture is infected with the virus. Serum can then be added back to the culture. If some other method is used to inactivate the factor, it can be discontinued, inactivated or removed (such as removing the anti-factor antibody, *e.g.*, with a bound antibody directed against that antibody) prior to adding fresh serum back to the culture. Cells that survive are mutants having an inactivating insertion in a gene necessary for growth of the virus. The genes having the insertions can then be isolated by isolating sequences having the marker gene sequences. This mutational process

disturbs a wild type function. A mutant gene may produce at a lower level a normal product, it may produce a normal product not normally found in these cells, it may cause the overproduction of a normal product, it may produce an altered product that has some functions but not others, or it may completely disrupt a gene function. Additionally, the mutation may disrupt an RNA that has a function but is never translated into a protein. For example, the alpha-tropomyosin gene has a 3' RNA that is very important in cell regulation but never is translated into protein. (*Cell* 75 pg 1107-1117, 12/17/93).

As used herein, a cellular gene "nonessential for cellular survival" means a gene for which disruption of one or both alleles results in a cell viable for at least a period of time which allows viral replication to be inhibited for preventative or therapeutic uses or use in research. A gene "necessary for viral growth" means the gene product, either protein or RNA, secreted or not, is necessary or beneficial, either directly or indirectly in some way for the virus to grow, and therefore, in the absence of that gene product (*i.e.*, a functionally available gene product), the virus does not spread. For example, such genes can encode cell cycle regulatory proteins, proteins affecting the vacuolar hydrogen pump, or proteins involved in protein folding and protein modification, including but not limited to: phosphorylation, methylation, glycosylation, myristylation or other lipid moiety, or protein processing via enzymatic processing. Some examples of such genes include vacuolar H<sup>+</sup>ATPase, alpha tropomyosin, gas5 gene, ras complex, N-acetylglucosaminy-l-transferase I mRNA, annexin II, c-golgi CM130 and calcyclin.

Any virus capable of infecting the cell can be used for this method. Virus can be selected based upon the particular infection desired to study. However, it is contemplated by the present invention that many viruses will be dependent upon the same cellular genes for survival; thus a cellular gene isolated using one virus can be used as a target for therapy for other viruses as well. Any cellular gene can be tested for relevancy to any desired virus using the methods set forth herein, *i.e.*, in general, by inhibiting the gene or its gene product in a cell and determining if the desired virus can grow in that cell. Some examples of viruses include HIV (including HIV-1 and HIV-2); parvovirus; papillomaviruses; hantaviruses; influenza viruses (*e.g.*, influenza A, B and C viruses); hepatitis viruses A to G; caliciviruses; astroviruses; rotaviruses; coronaviruses, such as human respiratory coronavirus; picornaviruses, such as human rhinovirus and enterovirus; ebola virus; human herpesvirus (*e.g.*, HSV-1-9); human adenovirus; for animal, the animal counterpart to any above listed human virus, animal retroviruses, such



as simian immunodeficiency virus, avian immunodeficiency virus, bovine immunodeficiency virus, feline immunodeficiency virus, equine infectious anemia virus, caprine arthritis encephalitis virus, arenaviruses, arvoviruses, tickborne viruses or visna virus.

5           The nucleic acids comprising cellular genes of this invention were isolated by the above method and as set forth in the examples. The invention includes a nucleic acid comprising the nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, 10 SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, 15 SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, 20 SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:112, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, 25 SEQ ID NO:125, SEQ ID NO:126, and SEQ ID NO:127 (this list is sometimes referred to herein as "SEQ LIST 1" for brevity). Thus these nucleic acids can contain, in addition to the nucleotides set forth in each SEQ ID NO in the sequence listing, additional nucleotides at either end of the molecule. Such additional nucleotides can be added by 30 any standard method, as known in the art, such as recombinant methods and synthesis methods. Examples of such nucleic acids comprising the nucleotide sequence set forth in any entry of the sequence listing contemplated by this invention include, but are not limited to, for example, the nucleic acid placed into a vector; a nucleic acid having one or



more regulatory region (*e.g.*, promoter, enhancer, polyadenylation site) linked to it, particularly in functional manner, *i.e.* such that an mRNA or a protein can be produced; a nucleic acid including additional nucleic acids of the gene, such as a larger or even full length genomic fragment of the gene, a partial or full length cDNA, a partial or full length RNA. Making and/or isolating such larger nucleic acids is further described below and is well known and standard in the art.

Also provided in this invention are the double-stranded nucleic acids corresponding to the nucleic acid sequences set forth in SEQ ID 1 through SEQ ID 136, inclusive. It is recognized that "nucleic acid" as used herein, can refer to either or both strands of such double-stranded nucleic acids, such strands often referred to as the "positive" and "negative" strands. Either strand of such double-stranded nucleic acids may encode the polypeptides of this invention, and the coding sequences for such polypeptides may be translated in either direction along the strand. Examples of polypeptides encoded by either strand are disclosed herein.

The invention also provides a nucleic acid encoding the protein encoded by the gene comprising the nucleotide sequence set forth in any of the sequences listed in SEQ LIST 1, as well as allelic variants and homologs of each such gene. The gene is readily obtained using standard methods, as described below and as is known and standard in the art. The present invention also contemplates any unique fragment of these genes or of the nucleic acids set forth in any of the sequences listed in SEQ LIST 1. Examples of inventive fragments of the inventive genes can include the nucleic acids whose sequence is set forth in any of the sequences listed in SEQ LIST 1. To be unique, the fragment must be of sufficient size to distinguish it from other known sequences, most readily determined by comparing any nucleic acid fragment to the nucleotide sequences of nucleic acids in computer databases, such as GenBank. Such comparative searches are standard in the art. Typically, a unique fragment useful as a primer or probe will be at least about 20 to about 25 nucleotides in length, depending upon the specific nucleotide content of the sequence. Additionally, fragments can be, for example, at least about 30, 40, 50, 75, 100, 200 or 500 nucleotides in length. The nucleic acids can be single or double stranded, depending upon the purpose for which it is intended.

The present invention further provides a nucleic acid comprising the regulatory region of a gene comprising any one of the nucleotide sequences set forth in SEQ LIST 1, as well as homologs of each such gene. Additionally provided is a construct

comprising such a regulatory region functionally linked to a reporter gene. Such reporter gene constructs can be used to screen for compounds and compositions that affect expression of the gene comprising the nucleic acids whose sequence is set forth in SEQ LIST 1, or any homologs thereof.

- 5           The nucleic acids set forth in the sequence listing are gene fragments; the entire coding sequence and the entire gene that comprises each fragment are both contemplated herein and are readily obtained by standard methods, given the nucleotide sequences presented in the sequence listing (*see, e.g.*, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989; *DNA cloning: A Practical Approach*, Volumes I and II, Glover, D.M. ed., IRL Press Limited, Oxford, 1985). To obtain the entire genomic gene, briefly, a nucleic acid whose sequence is set forth in any of SEQ ID NO:1 through SEQ ID NO:127, or preferably in any of the sequences listed in SEQ LIST 1, or a smaller fragment thereof, is utilized as a probe to screen a genomic library under high stringency conditions, and
- 10           isolated clones are sequenced. Once the sequence of the new clone is determined, a probe can be devised from a portion of the new clone not present in the previous fragment and hybridized to the library to isolate more clones containing fragments of the gene. In this manner, by repeating this process in organized fashion, one can "walk" along the chromosome and eventually obtain nucleotide sequence for the entire gene.
- 15           Similarly, one can use portions of the present fragments, or additional fragments obtained from the genomic library, that contain open reading frames to screen a cDNA library to obtain a cDNA having the entire coding sequence of the gene. Repeated screens can be utilized as described above to obtain the complete sequence from several clones if necessary. The isolates can then be sequenced to determine the nucleotide
- 20           sequence by standard means such as dideoxynucleotide sequencing methods (*see, e.g.*, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989).

- 25           The present genes were isolated from rat; however, homologs in any desired species, preferably mammalian, such as human, can readily be obtained by screening a
- 30           human library, genomic or cDNA, with a probe comprising sequences of the nucleic acids set forth in the sequence listing herein, or fragments thereof, and isolating genes specifically hybridizing with the probe under preferably relatively high stringency hybridization conditions. For example, high salt conditions (*e.g.*, in 6X SSC or 6X

SSPE) and/or high temperatures of hybridization can be used. For example, the stringency of hybridization is typically about 5°C to 20°C below the  $T_m$  (the melting temperature at which half of the molecules dissociate from its partner) for the given chain length. As is known in the art, the nucleotide composition of the hybridizing region factors in determining the melting temperature of the hybrid. For 20mer probes, for example, the recommended hybridization temperature is typically about 55-58°C. Additionally, the rat sequence can be utilized to devise a probe for a homolog in any specific animal by determining the amino acid sequence for a portion of the rat protein, and selecting a probe with optimized codon usage to encode the amino acid sequence of the homolog in that particular animal. Any isolated gene can be confirmed as the targeted gene by sequencing the gene to determine it contains the nucleotide sequence listed herein as comprising the gene. Any homolog can be confirmed as a homolog by its functionality.

Additionally contemplated by the present invention are nucleic acids, from any desired species, preferably mammalian and more preferably human, having 98%, 95%, 90%, 85%, 80%, 70%, 60%, or 50% homology, or greater, in the region of homology, to a region in an exon of a nucleic acid encoding the protein encoded by the gene comprising the nucleotide sequence set forth in any of the sequences listed in SEQ LIST 1 or to homologs thereof. Also contemplated by the present invention are nucleic acids, from any desired species, preferably mammalian and more preferably human, having 98%, 95%, 90%, 85%, 80%, 70%, 60%, or 50% homology, or greater, in the region of homology, to a region in an exon of a nucleic acid comprising the nucleotide sequence set forth in any of the sequences listed in SEQ LIST 1 or to homologs thereof. These genes can be synthesized or obtained by the same methods used to isolate homologs, with stringency of hybridization and washing, if desired, reduced accordingly as homology desired is decreased, and further, depending upon the G-C or A-T richness of any area wherein variability is searched for. Allelic variants of any of the present genes or of their homologs can readily be isolated and sequenced by screening additional libraries following the protocol above. Methods of making synthetic genes are described in U.S. Patent No. 5,503,995 and the references cited therein.

The nucleic acid encoding any selected protein of the present invention can be any nucleic acid that functionally encodes that protein. For example, to functionally encode, *i.e.*, allow the nucleic acid to be expressed, the nucleic acid can include, for

example, exogenous or endogenous expression control sequences, such as an origin of replication, a promoter, an enhancer, and necessary information processing sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences can be promoters derived  
5 from metallothionine genes, actin genes, immunoglobulin genes, CMV, SV40, adenovirus, bovine papilloma virus, etc. Expression control sequences can be selected for functionality in the cells in which the nucleic acid will be placed. A nucleic acid encoding a selected protein can readily be determined based upon the amino acid sequence of the selected protein, and, clearly, many nucleic acids will encode any selected  
10 protein.

The present invention additionally provides a nucleic acid that selectively hybridizes under stringent conditions with a nucleic acid set forth in SEQ LIST 1 or with a nucleic acid encoding the protein encoded by the gene comprising the nucleotide sequence set forth in any sequence listed in SEQ LIST 1. This hybridization can be  
15 specific. The degree of complementarity between the hybridizing nucleic acid and the sequence to which it hybridizes should be at least enough to exclude hybridization with a nucleic acid encoding an unrelated protein. Thus, a nucleic acid that selectively hybridizes with a nucleic acid of the present protein coding sequence will not selectively hybridize under stringent conditions with a nucleic acid for a different, unrelated protein,  
20 and vice versa. Typically, the stringency of hybridization to achieve selective hybridization involves hybridization in high ionic strength solution (6X SSC or 6X SSPE) at a temperature that is about 12-25°C below the  $T_m$  (the melting temperature at which half of the molecules dissociate from its partner) followed by washing at a combination of temperature and salt concentration chosen so that the washing temperature is about 5°C  
25 to 20°C below the  $T_m$  of the hybrid molecule. The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters are hybridized to a labeled nucleic acid of interest and then washed under conditions of different stringencies. Hybridization temperatures are typically higher for DNA-RNA and RNA-RNA hybridizations. The washing  
30 temperatures can be used as described above to achieve selective stringency, as is known in the art. (Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989; Kunkel et al. *Methods Enzymol.* 1987:154:367, 1987). Nucleic acid fragments that selectively hybridize to any

given nucleic acid can be used, *e.g.*, as primers and or probes for further hybridization or for amplification methods (*e.g.*, polymerase chain reaction (PCR), ligase chain reaction (LCR)). A preferable stringent hybridization condition for a DNA:DNA hybridization can be at about 68°C (in aqueous solution) in 6X SSC or 6X SSPE followed by washing  
 5 at 68°C.

The present invention additionally provides a polypeptide comprising the amino acid sequence encoded by the gene comprising the nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ  
 10 ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID  
 15 NO:45, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID  
 20 NO:80, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108,  
 25 SEQ ID NO:112, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, and SEQ ID NO:127 (*i.e.*, SEQ LIST 1). Additionally, polypeptides comprising the amino acid sequence encoded by a nucleic acid that selectively hybridizes under stringent conditions with a nucleic acid in SEQ LIST 1 are provided. Further, polypeptides comprising the amino acid sequence  
 30 encoded by a nucleic acid having a region within an exon wherein the region has at least 50, 60, 70, 80, 90, or 95% homology with a nucleic acid in SEQ LIST 1. These polypeptides can be readily obtained by any of several means. For example, the nucleotide sequence of coding regions of the gene can be translated and then the



corresponding polypeptide can be synthesized mechanically by standard methods. Additionally, the coding regions of the genes can be expressed or synthesized, an antibody specific for the resulting polypeptide can be raised by standard methods (see, e.g., Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1988), and the protein can be isolated from other cellular proteins by selective hybridization with the antibody. This protein can be purified to the extent desired by standard methods of protein purification (see, e.g., Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989). The amino acid sequence of any protein, polypeptide or peptide of this invention can be deduced from the nucleic acid sequence, or it can be determined by sequencing an isolated or recombinantly produced protein.

The terms "peptide," "polypeptide" and "protein" can be used interchangeably herein and refer to a polymer of amino acids and includes full-length proteins and fragments thereof. As used in the specification and in the claims, "a" can mean one or more, depending upon the context in which it is used. An amino acid residue is an amino acid formed upon chemical digestion (hydrolysis) of a polypeptide at its peptide linkages. The amino acid residues described herein are preferably in the L isomeric form. However, residues in the D isomeric form can be substituted for any L-amino acid residue, as long as the desired functional property is retained by the polypeptide. Standard polypeptide nomenclature (described in *J. Biol. Chem.*, 243:3552-59 (1969) and adopted at 37 CFR § 1.822(b)) is used herein.

As will be appreciated by those skilled in the art, the invention also includes those polypeptides having slight variations in amino acid sequences or other properties. Amino acid substitutions can be selected by known parameters to be neutral (see, e.g., Robinson WE Jr, and Mitchell WM., *AIDS* 4:S151-S162(1990)). Such variations may arise naturally as allelic variations (e.g., due to genetic polymorphism) or may be produced by human intervention (e.g., by mutagenesis of cloned DNA sequences), such as induced point, deletion, insertion and substitution mutants. Minor changes in amino acid sequence are generally preferred, such as conservative amino acid replacements, small internal deletions or insertions, and additions or deletions at the ends of the molecules. Substitutions may be designed based on, for example, the model of Dayhoff, *et al.* (in *Atlas of Protein Sequence and Structure* 1978, Nat'l Biomed. Res. Found., Washington,



D.C.). These modifications can result in changes in the amino acid sequence, provide silent mutations, modify a restriction site, or provide other specific mutations. Likewise, such amino acid changes result in a different nucleic acid encoding the polypeptides and proteins. Thus, alternative nucleic acids are also contemplated by such modifications.

5       The present invention also provides cells containing a nucleic acid of the invention. A cell containing a nucleic acid encoding a protein typically can replicate the DNA and, further, typically can express the encoded protein. The cell can be a prokaryotic cell, particularly for the purpose of producing quantities of the nucleic acid, or a eukaryotic cell, particularly a mammalian cell. The cell is preferably a mammalian  
10 cell for the purpose of expressing the encoded protein so that the resultant produced protein has mammalian protein processing modifications.

Nucleic acids of the present invention can be delivered into cells by any selected means, in particular depending upon the purpose of the delivery of the compound and the target cells. Many delivery means are well-known in the art. For example,  
15 electroporation, calcium phosphate precipitation, microinjection, cationic or anionic liposomes, and liposomes in combination with a nuclear localization signal peptide for delivery to the nucleus can be utilized, as is known in the art.

The present invention also contemplates that the mutated cellular genes necessary for viral growth, produced by the present method, as well as cells containing these  
20 mutants can also be useful. These mutated genes and cells containing them can be isolated and/or produced according to the methods herein described and using standard methods.

It should be recognized that the sequences set forth herein may contain minor sequencing errors. Such errors can be corrected, for example, by using the hybridization  
25 procedure described above with various probes derived from the described sequences such that the coding sequence can be reisolated and resequenced.

As described in the examples, the present invention provides the discovery of a "serum survival factor" present in serum that is necessary for the survival of persistently virally infected cells. Isolation and characterization of this factor have shown it to be a  
30 protein, to have a molecular weight of between about 50 kD and 100 kD, to resist inactivation in low pH (*e.g.*, pH2) and chloroform extraction, to be inactivated by boiling for about 5 minutes and in low ionic strength solution (*e.g.*, about 10 mM to about 50 mM). The present invention thus provides a purified mammalian serum protein having a

molecular weight of between about 50 kD and 100 kD which resists inactivation in low pH and resists inactivation by chloroform extraction, which inactivates when boiled and inactivates in low ionic strength solution, and which when removed from a cell culture comprising cells persistently infected with reovirus selectively substantially prevents survival of cells persistently infected with reovirus. The factor, fitting the physical characteristics described above, can readily be verified by adding it to non-serum-containing medium (which previously could not support survival of persistently virally infected cells) and determining whether this medium with the added putative factor can now support persistently virally infected cells, particularly cells persistently infected with reovirus. As used herein, a "purified" protein means the protein is at least of sufficient purity such that an approximate molecular weight can be determined.

The amino acid sequence of the protein can be elucidated by standard methods. For example, an antibody to the protein can be raised and used to screen an expression library to obtain nucleic acid sequence coding the protein. This nucleic acid sequence is then simply translated into the corresponding amino acid sequence. Alternatively, a portion of the protein can be directly sequenced by standard amino acid sequencing methods (amino-terminus sequencing). This amino acid sequence can then be used to generate an array of nucleic acid probes that encompasses all possible coding sequences for a portion of the amino acid sequence. The array of probes is used to screen a cDNA library to obtain the remainder of the coding sequence and thus ultimately the corresponding amino acid sequence.

The present invention also provides methods of detecting and isolating additional serum survival factors. For example, to determine if any known serum components are necessary for viral growth, the known components can be inhibited in, or eliminated from, the culture medium, and it can be observed whether viral growth is inhibited by determining if persistently infected cells do not survive. One can add the factor back (or remove the inhibition) and determine whether the factor allows for viral growth.

Additionally, other, unknown serum components can also be found to be essential for growth. Serum can be fractionated by various standard means, and fractions added to serum free medium to determine if a factor is present in a reaction that allows growth previously inhibited by the lack of serum. Fractions having this activity can then be further fractionated until the factor is relatively free of other components. The factor can then be characterized by standard methods, such as size fractionation, denaturation and/or

inactivation by various means, etc. Preferably, once the factor has been purified to a desired level of purity, it is added to cells in serum free medium to confirm that it bestows the function of allowing virus to grow when serum-free medium alone did not. This method can be repeated to confirm the requirement for the specific factor for any desired virus, since each serum factor found to be required by any one virus can also be required by many other viruses. In general, the closer the viruses are related and the more similar the infection modes of the viruses, the more likely that a factor required by one virus will be required by the other.

The present invention also provides methods of treating virus infections utilizing applicants' discoveries. The subject of any of the herein described methods can be any animal, preferably a mammal, such as a human, a veterinary animal, such as a cat, dog, horse, pig, goat, sheep, or cow, or a laboratory animal, such as a mouse, rat, rabbit, or guinea pig, depending upon the virus.

The present invention provides a method of reducing or inhibiting, and thereby treating, a viral infection in a subject, comprising administering to the subject an inhibiting amount of a composition that inhibits functioning of the serum protein described herein, *i.e.* the serum protein having a molecular weight of between about 50 kD and 100 kD which resists inactivation in low pH and resists inactivation by chloroform extraction, which inactivates when boiled and inactivates in low ionic strength solution, and which when removed from a cell culture comprising cells persistently infected with the virus prevents survival of at least some cells persistently infected with the virus, thereby treating the viral infection. The composition can comprise, for example, an antibody that specifically binds the serum protein, or an antisense RNA that binds an RNA encoded by a gene functionally encoding the serum protein.

Any virus capable of infecting the selected subject to be treated can be treated by the present methods. As described above, any serum protein or survival factor found by the present methods to be necessary for growth of cells infected with any one virus can be found to be necessary for growth of the cells infected with many other viruses. For any given cell-virus combination, the serum protein or factor can be confirmed to be required for growth by the methods described herein. The cellular genes identified by the examples using reovirus, a mammalian pathogen, and a rat cell system have general applicability to other virus infections that include all of the known as well as yet to be discovered human pathogens, including, but not limited to: human immunodeficiency

viruses (e.g., HIV-1, HIV-2); parvovirus; papillomaviruses; hantaviruses; influenza viruses (e.g., influenza A, B and C viruses); hepatitis viruses A to G; caliciviruses; astroviruses; rotaviruses; coronaviruses, such as human respiratory coronavirus; picornaviruses, such as human rhinovirus and enterovirus; ebola virus; human herpesvirus (e.g., HSV-1-9); human adenovirus; hantaviruses; for animal, the animal counterpart to any above listed human virus, animal retroviruses, such as simian immunodeficiency virus, avian immunodeficiency virus, bovine immunodeficiency virus, feline immunodeficiency virus, equine infectious anemia virus, caprine arthritis encephalitis virus, arenaviruses, arvoviruses, tickborne virus or visna virus.

10 A protein inhibiting amount of the composition can be readily determined, such as by administering varying amounts to cells or to a subject and then adjusting the effective amount for inhibiting the protein according to the volume of blood or weight of the subject. Compositions that bind to the protein can be readily determined by running the putatively bound protein on a protein gel and observing an alteration in the protein's  
15 migration through the gel. Inhibition of the protein can be determined by any desired means such as adding the inhibitor to complete media used to maintain persistently infected cells and observing the cells' viability. The composition can comprise, for example, an antibody that specifically binds the serum protein. Specific binding by an antibody means that the antibody can be used to selectively remove the factor from serum  
20 or inhibit the factor's biological activity and can readily be determined by radio immune assay (RIA), bioassay, or enzyme-linked immunosorbant (ELISA) technology. The composition can comprise, for example, an antisense RNA that specifically binds an RNA encoded by the gene encoding the serum protein. Antisense RNAs can be synthesized and used by standard methods (e.g., *Antisense RNA and DNA*, D. A. Melton, Ed., Cold  
25 Spring Harbor Laboratory, Cold Spring Harbor, NY (1988)).

The present methods provide a method of screening a compound for effectiveness in treating or preventing a viral infection, comprising administering the compound to a cell containing a cellular gene functionally encoding a gene product necessary for reproduction of the virus in the cell but not necessary for survival of the cell and detecting the level  
30 and/or activity (i.e. function) of the gene product produced, a decrease or elimination of the gene product and/or the gene product activity indicating a compound for treating or preventing the viral infection. The cellular gene can be, for example, a nucleic acid set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ

ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:47, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:60, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:72, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:75, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:90, SEQ ID NO:91, SEQ ID NO:92, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, or SEQ ID NO:127 (herein sometimes referred to as SEQ LIST 2, for brevity), any homolog thereof, or any other gene obtained using the methods provided herein for obtaining such genes. It is understood that the cellular gene can be present naturally in the cell being screened, or it can be introduced into the cell in a suitable expression vector, as are well known in the art. The level of the gene product can be measured by any standard means, such as by detection with an antibody specific for the protein. The level of gene product can be compared to the level of the gene product in a control cell not contacted with the compound. The level of gene product can be compared to the level of the gene product in the same cell prior to addition of the compound. Activity, or function, can be measured by any standard means, such as by enzymatic assays that measure the conversion of a substrate to a product or binding



assays that measure the binding of a protein to a nucleic acid, for example. Examples of gene products disclosed herein whose activity/function can be measured include tristetraprolin (human ZFP-36), 6-pyruvoyl-tetrahydropterin synthase, a eukaryotic DnaJ-like protein, ID3 (inhibitor of DNA binding 3), N-acetylglucos-aminyltransferase I (mGAT-1), cleavage stimulation factor (CSTF2), TAK1 binding protein, human zinc transcription factor ZPF207, Dlx2, Smad7 (Mad-related protein), and P-glycoprotein (mdr1b). The activity can be compared to the activity in a control cell not contacted with the compound or in the same cell prior to addition of the compound. Relatedly, the regulatory region of the gene can be functionally linked to a reporter gene and compounds can be screened for inhibition of the reporter gene. Such reporter constructs are described herein.

The present invention also provides a method of screening a compound for effectiveness in treating or preventing a viral infection comprising contacting the compound with the gene product of a cellular gene comprising a nucleic acid of SEQ LIST 2, or any homolog thereof, and detecting the function of the gene product, a decrease or elimination of the function indicating a compound effective for treating or preventing viral infection. Examples of gene products disclosed herein that can be utilized in this method include tristetraprolin (human ZFP-36), 6-pyruvoyl-tetrahydropterin synthase, a eukaryotic DnaJ-like protein, ID3 (inhibitor of DNA binding 3), N-acetylglucos-aminyltransferase I (mGAT-1), cleavage stimulation factor (CSTF2), TAK1 binding protein, human zinc transcription factor ZPF207, Dlx2, Smad7 (Mad-related protein), and P-glycoprotein (mdr1b).

The present invention provides a method of selectively eliminating cells persistently infected with a virus from an animal cell culture capable of surviving for a first period of time in the absence of serum, comprising propagating the cell culture in the absence of serum for a second time period during which a persistently infected cell cannot survive without serum, thereby selectively eliminating from the cell culture cells persistently infected with the virus. The second time period should be shorter than the first time period. Thus one can simply eliminate serum from a standard culture medium composition for a period of time (*e.g.* by removing serum containing medium from the culture container, rinsing the cells, and adding serum-free medium back to the container), then, after a time of serum starvation, return serum to the culture medium. Alternatively, one can inhibit a serum survival factor from the culture in place of the step of serum starvation. Furthermore, one can instead interfere with the virus-factor interaction. Such a



viral elimination method can periodically be performed for cultured cells to ensure that they remain virus-free. The time period of serum removal can greatly vary, with a typical range being about 1 to about 30 days; a preferable period can be about 3 to about 10 days, and a more preferable period can be about 5 days to about 7 days. This time period can be  
5 selected based upon ability of a specific cell to survive without serum as well as the life cycle of the target virus, *e.g.*, for reovirus, which has a life cycle of about 24 hours, 3 days' starvation of cells provides dramatic results.

Furthermore, the time period can be shortened by also passaging the cells during the starvation; in general, increasing the number of passages can decrease the time of  
10 serum starvation (or serum factor inhibition) needed to get full clearance of the virus from the culture. While passaging, the cells typically are exposed briefly to serum (typically for about 3 to about 24 hours). This exposure both stops the action of the trypsin used to dislodge the cells and stimulates the cells into another cycle of growth, thus aiding in this selection process. Thus a starvation/serum cycle can be repeated to optimize the selective  
15 effect. Other standard culture parameters, such as confluency of the cultures, pH, temperature, etc. can be varied to alter the needed time period of serum starvation (or serum survival factor inhibition). This time period can readily be determined for any given viral infection by simply removing the serum for various periods of time, then testing the cultures for the presence of the infected cells (*e.g.*, by ability to survive in the  
20 absence of serum and confirmed by quantitating virus in cells by standard virus titration and immunohistochemical techniques) at each tested time period, and then detecting at which time periods of serum deprivation the virally infected cells were eliminated. It is preferable that shorter time periods of serum deprivation that still provide elimination of the persistently infected cells be used. Furthermore, the cycle of starvation, then adding  
25 back serum and determining amount of virus remaining in the culture can be repeated until no virtually infected cells remain in the culture.

Thus, the present method can further comprise passaging the cells, *i.e.*, transferring the cell culture from a first container to a second container. Such transfer can facilitate the selective lack of survival of virally infected cells. Transfer can be repeated several times.  
30 Transfer is achieved by standard methods of tissue culture (*see, e.g.*, Freshney, *Culture of Animal Cells, A Manual of Basic Technique*, 2nd Ed. Alan R. Liss, Inc., New York, 1987).

The present method further provides a method of selectively eliminating from a cell culture cells persistently infected with a virus, comprising propagating the cell culture

in the absence of a functional form of the serum protein having a molecular weight of between about 50 kD and 100 kD which resists inactivation in low pH and resists inactivation by chloroform extraction, which inactivates when boiled and inactivates in low ionic strength solution, and which when removed from a cell culture comprising cells  
5 persistently infected with reovirus substantially prevents survival of cells persistently infected with reovirus. The absence of the functional form can be achieved by any of several standard means, such as by binding the protein to an antibody selective for it (binding the antibody in serum either before or after the serum is added to the cells; if before, the serum protein can be removed from the serum by, *e.g.*, binding the antibody to  
10 a column and passing the serum over the column and then administering the survival protein-free serum to the cells), by administering a compound that inactivates the protein, or by administering a compound that interferes with the interaction between the virus and the protein.

Thus, the present invention provides a method of selectively eliminating from a  
15 cell culture propagated in serum-containing medium cells persistently infected with a virus, comprising inhibiting in the serum the protein having a molecular weight of between about 50 kD and 100 kD which resists inactivation in low pH and resists inactivation by chloroform extraction, which inactivates when boiled and inactivates in low ionic strength solution, and which when removed from a cell culture comprising cells persistently  
20 infected with reovirus substantially prevents survival of cells persistently infected with reovirus. Alternatively, the interaction between the virus and the serum protein can be disrupted to selectively eliminate cells persistently infected with the virus.

Any virus capable of some form of persistent infection may be eliminated from a cell culture utilizing the present elimination methods, including removing, inhibiting or  
25 otherwise interfering with a serum protein, such as the one exemplified herein, and also including removing, inhibiting or otherwise interfering with a gene product from any cellular gene found by the present method to be necessary for viral growth yet nonessential to the cell. For example, DNA viruses or RNA viruses can be targeted. One can readily determine whether cells infected with a selected virus can be selectively removed from a  
30 culture through removal of serum by starving cells permissive to the virus of serum (or inhibiting the serum survival factor), adding the selected virus to the cells, adding serum to the culture, and observing whether infected cells die (*i.e.*, by titering levels of virus in the surviving cells with an antibody specific for the virus).

A culture of any animal cell (*i.e.*, any cell that is typically grown and maintained in culture in serum) that can be maintained for a period of time in the absence of serum, can be purified from viral infection utilizing the present method. For example, primary cultures as well as established cultures and cell lines can be used. Furthermore, cultures of cells from any animal and any tissue or cell type within that animal that can be cultured and that can be maintained for a period of time in the absence of serum can be used. For example, cultures of cells from tissues typically infected, and particularly persistently infected, by an infectious virus could be used.

As used in the claims "in the absence of serum" means at a level at which persistently virally infected cells do not survive. Typically, the threshold level is about 1% serum in the media. Therefore, about 1% serum or less can be used, such as about 1%, 0.75%, 0.50%, 0.25%, 0.1% or no serum can be used.

As used herein, "selectively eliminating" cells persistently infected with a virus means that substantially all of the cells persistently infected with the virus are killed such that the presence of virally infected cells cannot be detected in the culture immediately after the elimination procedure has been performed. Furthermore, "selectively eliminating" includes that cells not infected with the virus are generally not killed by the method. Some surviving cells may still produce virus but at a lower level, and some may be defective in pathways that lead to death by the virus. Typically, for cells persistently infected with virus to be substantially all killed, more than about 90% of the cells, and more preferably more than about 95%, 98%, 99%, or 99.99% of virus-containing cells in the culture are killed.

The present method also provides a nucleic acid comprising the regulatory region of any of the genes. Such regulatory regions can be isolated from the genomic sequences isolated and sequenced as described above and identified by any characteristics observed that are characteristic for regulatory regions of the species and by their relation to the start codon for the coding region of the gene. The present invention also provides a construct comprising the regulatory region functionally linked to a reporter gene. Such constructs are made by routine subcloning methods, and many vectors are available into which regulatory regions can be subcloned upstream of a marker gene. Marker genes can be chosen for ease of detection of marker gene product.

The present method therefore also provides a method of screening a compound for treating a viral infection, comprising administering the compound to a cell containing any

of the above-described constructs, comprising a regulatory region of one of the genes comprising any of the nucleotide sequences set forth in SEQ LIST 2, or any homologs thereof, whose inhibition or reduction in expression causes inhibition of viral replication wherein the region is functionally linked to a reporter gene, and detecting the level of the reporter gene product produced, a decrease or elimination of the reporter gene product indicating a compound for treating the viral infection. Compounds detected by this method would inhibit transcription of the gene from which the regulatory region was isolated, and thus, in treating a subject, would inhibit the production of the gene product produced by the gene, and thus treat the viral infection.

Some genes when disrupted by the present method of retrovirus insertion, resulted in over expression of the gene product, and this overexpression inhibited viral replication. Thus the present invention provides a method of screening a compound for effectiveness in treating a viral infection, comprising administering the compound to a cell containing a cellular gene functionally encoding a gene product whose overexpression inhibits reproduction of the virus but does not prevent survival of the cell and detecting the level of the gene product produced, an increase in the gene product indicating a compound effective for treating the viral infection. Typically, an increase will be a 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 125%, 150%, 175%, 200%, 300%, 400%, 500% or higher increase over gene product produced when the compound is not present.

The present invention additionally provides a method of reducing or inhibiting a viral infection in a subject, comprising administering to the subject an amount of a composition that inhibits expression or functioning of a gene product encoded by a gene comprising the nucleic acid set forth in any of SEQ LIST 2, or a homolog thereof, thereby treating the viral infection. Reducing or inhibiting viral infection naturally can include both the initial infection of the subject and the infection of uninfected cells within an already infected subject, e.g. inhibiting viral replication in cells of the subject. The composition can comprise, for example, an antibody that binds a protein encoded by the gene. The composition can also comprise an antibody that binds a receptor for a protein encoded by the gene. Such an antibody can be raised against the selected protein by standard methods as set forth above, and can be either polyclonal or monoclonal, though monoclonal is preferred. Alternatively, the composition can comprise an antisense RNA that binds an RNA encoded by the gene, as described above. Examples of antisense RNA useful therapeutically include the fragments of the nucleic acids described above.

Furthermore, the composition can comprise a nucleic acid functionally encoding an antisense RNA that binds an RNA encoded by the gene. Other useful compositions will be readily apparent to the skilled artisan.

The present invention also provides a method of treating a viral infection in a subject comprising administering to the subject a treatment effective amount of a composition that increases expression of a gene whose over expression reduces or inhibits viral replication. Typically, an increase will be a 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 125%, 150%, 175%, 200%, 300%, 400%, 500% or higher increase over gene product produced when the composition is not present.

The present invention further provides a method of reducing or inhibiting a viral infection in a subject comprising mutating *ex vivo* in a selected cell, for example from the subject or from an allogenic source, an endogenous gene comprising a nucleic acid set forth in SEQ LIST 2 whose inhibition or reduction in expression causes inhibition of viral replication, or a homolog thereof, to a gene form incapable of producing a functional gene product of the gene or a gene form producing a reduced amount of a functional gene product of the gene, and placing (or replacing, in the case of the subject's own cells) the cell in the subject, thereby reducing viral infection of cells in the subject. The cell can be selected according to the typical target cell of the specific virus whose infection is to be reduced, prevented or inhibited. A preferred cell for several viruses is a hematopoietic cell. When the selected cell is a hematopoietic cell, viruses which can be reduced or inhibited from infection can include, for example, HIV, including HIV-1 and HIV-2. However, many other virus-cell combinations will be apparent to the skilled artisan.

The invention also includes a method of reducing or inhibiting viral infection in a subject comprising mutating *ex vivo* in a selected cell, for example from a subject or an allogenic source, an endogenous gene comprising a nucleic acid set forth in SEQ LIST 2 whose overexpression causes inhibition of viral replication, or a homolog thereof, to a gene form that expresses the gene at a higher level than the endogenous gene, and placing or replacing the cell in the subject. Typically, a higher level can be 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 125%, 150%, 175%, 200%, 300%, 400%, 500% or higher than the non-mutated, endogenous gene. The cell can be selected according to the typical target cell of the specific virus whose infection is to be reduced, prevented or inhibited. A preferred cell for several viruses is a hematopoietic cell. When the selected cell is a hematopoietic cell, viruses which can be reduced or inhibited from infection can



include, for example, HIV, including HIV-1 and HIV-2. However, many other virus-cell combinations will be apparent to the skilled artisan.

The present invention additionally provides a method of increasing viral infection resistance in a subject comprising mutating *ex vivo* in a selected cell, for example from the  
5 subject or from an allogenic source, an endogenous gene comprising a nucleic acid set forth in SEQ LIST 2, whose inhibition or reduction in expression increases viral infection resistance, said endogenous gene being mutated to a mutated gene form incapable of producing a functional gene product of the gene or a gene form producing a reduced amount of a functional gene product of the gene, and placing the cell in the subject,  
10 thereby increasing viral infection resistance of cells in the subject. The virus can be HIV, particularly when the cell is a hematopoietic cell. However, many other virus-cell combinations will be apparent to the skilled artisan.

Furthermore, the present invention provides a method for isolation of cellular genes utilized in tumor progression. The present invention provides a method of identifying a  
15 cellular gene that can suppress a malignant phenotype in a cell, comprising (a) transferring into a cell culture incapable of growing well in soft agar or Matrigel a vector encoding a selective marker gene lacking a functional promoter, (b) selecting cells expressing the marker gene, and (c) isolating from selected cells which are capable of growing in soft agar or Matrigel a cellular gene within which the marker gene is inserted, thereby  
20 identifying a gene that can suppress a malignant phenotype in a cell. This method can be performed using any selected non-transformed cell line, of which many are known in the art.

The present invention additionally provides a method of identifying a cellular gene that can suppress a malignant phenotype in a cell, comprising (a) transferring into a cell  
25 culture of non-transformed cells a vector encoding a selective marker gene lacking a functional promoter, (b) selecting cells expressing the marker gene, and (c) isolating from selected and transformed cells a cellular gene within which the marker gene is inserted, thereby identifying a gene that can suppress a malignant phenotype in a cell. A non-transformed phenotype can be determined by any of several standard methods in the  
30 art, such as the exemplified inability to grow in soft agar, or inability to grow in Matrigel.

The present invention further provides a method of screening for a compound for suppressing a malignant phenotype in a cell comprising administering the compound to a cell containing a cellular gene functionally encoding a gene product involved in

establishment of a malignant phenotype in the cell and detecting the level of the gene product produced, a decrease, inhibition or elimination of the gene product indicating a compound effective for suppressing the malignant phenotype. Detection of the level, or amount, of gene product produced can be measured, directly or indirectly, by any of several methods standard in the art (*e.g.*, protein gel, antibody-based assay, detecting labeled RNA) for assaying protein levels or amounts, and selected based upon the specific gene product.

The present invention also provides a method of screening for a compound for suppressing a malignant phenotype in a cell comprising administering the compound to a cell containing a cellular gene functionally encoding a gene product whose overexpression is involved in suppressing a malignant phenotype in the cell and detecting the level of the gene product produced, an increase in the gene product indicating a compound effective for suppressing the malignant phenotype.

The present invention further provides a method of suppressing a malignant phenotype in a cell in a subject, comprising administering to the subject an amount of a composition that inhibits expression or functioning of a gene product encoded by a gene comprising the nucleic acid set forth in SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, SEQ ID NO:36 or SEQ ID NO:94, or a homolog thereof, or any gene whose overexpression is found by the present method to be involved in suppressing a malignant phenotype in the cell (*e.g.*, any clone designated herein with an "x") thereby suppressing a malignant phenotype. The composition can, for example, comprise an antibody that binds a protein encoded by the gene. The composition can, as another example, comprise an antibody that binds a receptor for a protein encoded by the gene. The composition can comprise an antisense RNA that binds an RNA encoded by the gene. Further, the composition can comprise a nucleic acid functionally encoding an antisense RNA that binds an RNA encoded by the gene.

The present invention further provides a method of suppressing a malignant phenotype in a cell in a subject, comprising administering to the subject an amount of a composition that increases expression of a gene product whose overexpression is involved in suppressing a malignant phenotype in the cell. The gene product can be the product of a gene wherein disruption of an upstream gene by the present vector resulted in overexpression of the downstream gene, and the overexpression of the downstream gene



demonstrated a transformed phenotype. The composition can be, for example, an inhibitor, such as a small molecule inhibitor, of the COX 2 enzyme.

Diagnostic or therapeutic agents of the present invention can be administered to a subject or an animal model by any of many standard means for administering therapeutics or diagnostics to that selected site or standard for administering that type of functional entity. For example, an agent can be administered orally, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, topically, transdermally, or the like. Agents can be administered, e.g., as a complex with cationic liposomes, or encapsulated in anionic liposomes. Compositions can include various amounts of the selected agent in combination with a pharmaceutically acceptable carrier and, in addition, if desired, may include other medicinal agents, pharmaceutical agents, carriers, adjuvants, diluents, etc. Parental administration, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution or suspension in liquid prior to injection, or as emulsions. Depending upon the mode of administration, the agent can be optimized to avoid degradation in the subject, such as by encapsulation, etc.

Dosages will depend upon the mode of administration, the disease or condition to be treated, and the individual subject's condition, but will be that dosage typical for and used in administration of antiviral or anticancer agents. Dosages will also depend upon the composition being administered, e.g., a protein or a nucleic acid. Such dosages are known in the art. Furthermore, the dosage can be adjusted according to the typical dosage for the specific disease or condition to be treated. Furthermore, viral titers in culture cells of the target cell type can be used to optimize the dosage for the target cells *in vivo*, and transformation from varying dosages achieved in culture cells of the same type as the target cell type can be monitored. Often a single dose can be sufficient; however, the dose can be repeated if desirable. The dosage should not be so large as to cause adverse side effects. Generally, the dosage will vary with the age, condition, sex and extent of the disease in the patient and can be determined by one of skill in the art. The dosage can also be adjusted by the individual physician in the event of any complication.

For administration to a cell in a subject, the composition, once in the subject, will of course adjust to the subject's body temperature. For *ex vivo* administration, the composition can be administered by any standard methods that would maintain viability of the cells, such as by adding it to culture medium (appropriate for the target cells) and

adding this medium directly to the cells. As is known in the art, any medium used in this method can be aqueous and non-toxic so as not to render the cells non-viable. In addition, it can contain standard nutrients for maintaining viability of cells, if desired. For *in vivo* administration, the complex can be added to, for example, a blood sample or a tissue sample from the patient, or to a pharmaceutically acceptable carrier, e.g., saline and buffered saline, and administered by any of several means known in the art. Examples of administration include parenteral administration, e.g., by intravenous injection including regional perfusion through a blood vessel supplying the tissues(s) or organ(s) having the target cell(s), or by inhalation of an aerosol, subcutaneous or intramuscular injection, topical administration such as to skin wounds and lesions, direct transfection into, e.g., bone marrow cells prepared for transplantation and subsequent transplantation into the subject, and direct transfection into an organ that is subsequently transplanted into the subject. Further administration methods include oral administration, particularly when the composition is encapsulated, or rectal administration, particularly when the composition is in suppository form. A pharmaceutically acceptable carrier includes any material that is not biologically or otherwise undesirable, i.e., the material may be administered to an individual along with the selected complex without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained.

Specifically, if a particular cell type *in vivo* is to be targeted, for example, by regional perfusion of an organ or tumor, cells from the target tissue can be biopsied and optimal dosages for import of the complex into that tissue can be determined *in vitro*, as described herein and as known in the art, to optimize the *in vivo* dosage, including concentration and time length. Alternatively, cultured cells of the same cell type can also be used to optimize the dosage for the target cells *in vivo*.

For either *ex vivo* or *in vivo* use, the complex can be administered at any effective concentration. An effective concentration is that amount that results in reduction, inhibition or prevention of the viral infection or in reduction or inhibition of the transformed phenotype of the cells.

A nucleic acid can be administered in any of several means, which can be selected according to the vector utilized, the organ or tissue, if any, to be targeted, and the characteristics of the subject. The nucleic acids, if desired in a pharmaceutically acceptable carrier such as physiological saline, can be administered systemically, such as

intravenously, intraarterially, orally, parenterally, subcutaneously. The nucleic acids can also be administered by direct injection into an organ or by injection into the blood vessel supplying a target tissue. For an infection of cells of the lungs or trachea, it can be administered intratracheally. The nucleic acids can additionally be administered topically, transdermally, etc.

The nucleic acid or protein can be administered in a composition. For example, the composition can comprise other medicinal agents, pharmaceutical agents, carriers, adjuvants, diluents, etc. Furthermore, the composition can comprise, in addition to the vector, lipids such as liposomes, such as cationic liposomes (e.g., DOTMA, DOPE, DC-cholesterol) or anionic liposomes. Liposomes can further comprise proteins to facilitate targeting a particular cell, if desired. Administration of a composition comprising a vector and a cationic liposome can be administered to the blood afferent to a target organ or inhaled into the respiratory tract to target cells of the respiratory tract. Regarding liposomes, see, e.g., Brigham et al. *Am. J. Resp. Cell. Mol. Biol.* 1:95-100 (1989); Felgner et al. *Proc. Natl. Acad. Sci USA* 84:7413-7417 (1987); U.S. Pat. No.4,897,355.

For a viral vector comprising a nucleic acid, the composition can comprise a pharmaceutically acceptable carrier such as phosphate buffered saline or saline. The viral vector can be selected according to the target cell, as known in the art. For example, adenoviral vectors, in particular replication-deficient adenoviral vectors, can be utilized to target any of a number of cells, because of its broad host range. Many other viral vectors are available, and their target cells are known.

Throughout this application, various publications are referenced. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

## EXAMPLES

30

### Selective elimination of virally infected cells from a cell culture

Rat intestinal cell line-1 cells (RIE-1 cells) were standardly grown in Dulbecco's modified Eagle's medium, high glucose, supplemented with 10% fetal bovine serum. To

begin the experiment, cells persistently infected with reovirus were grown to near confluence, then serum was removed from the growth medium by removing the medium, washing the cells in PBS, and returning to the flask medium not supplemented with serum. Typically, the serum content was reduced to 1% or less. The cells are starved for serum  
5 for several days, or as long as about a month, to bring them to quiescence or growth arrest. Media containing 10% serum is then added to the quiescent cells to stimulate growth of the cells. Surviving cells are found to not be persistently infected cells by immunohistochemical techniques used to establish whether cells contain any infectious virus (sensitivity to 1 infectious virus per ml of homogenized cells).

10

#### **Cellular Genomic DNA Isolation**

Gene Trap Libraries: The libraries are generated by infecting the RIE-1 cells with a retrovirus vector (U3 gene-trap) at a ratio of less than one retrovirus for every ten cells. When a U3 gene trap retrovirus integrates within an actively transcribed gene, the  
15 neomycin resistance gene that the U3 gene trap retrovirus encodes is also transcribed, thus conferring resistance to the cell to the antibiotic neomycin. Cells with gene trap events are able to survive exposure to neomycin while cells without a gene trap event die. The various cells that survive neomycin selection are then propagated as a library of gene trap events. Such libraries can be generated with any retrovirus vector that has the properties  
20 of expressing a reporter gene from a transcriptionally active cellular promoter that tags the gene for later identification.

Reovirus selection: Reovirus infection is typically lethal to RIE-1 cells but can result in the development of persistently infected cells. These cells continue to grow while producing infective reovirus particles. For the identification of gene trap events that  
25 confer reovirus resistance to cells, the persistently infected cells must be eliminated or they will be scored as false positives. We have found that RIE-1 cells persistently infected with reovirus are very poorly tolerant to serum starvation, passaging and plating at low density. Thus, we have developed protocols for the screening of the RIE-1 gene trap libraries that select against both reovirus sensitive cells and cells that are persistently infected with  
30 reovirus.

1. RIE-1 library cells are grown to near confluence and then the serum is removed from the media. The cells are starved for serum for several days to bring them to quiescent or growth arrest.

2. The library cells are infected with reovirus at a titer of greater than ten reovirus per cell and the serum starvation is continued for several more days.
3. The infected cells are passaged, (a process in which they are exposed to serum for three to six hours) and then starved for serum for several more days.
- 5 4. The surviving cells are then allowed to grow in the presence of serum until visible colonies develop at which point they are cloned by limiting dilution.

MEDIA: DULBECCO'S MODIFIED EAGLE'S MEDIUM, HIGH GLUCOSE (DME/HIGH) Hyclone Laboratories cat. no. SH30003.02.

NEOMYCIN: The antibiotic used to select against the cells that did not have a U3 gene trap retrovirus, e.g. GENETICIN, from Sigma. [cat. no. G9516].

RAT INTESTINAL CELL LINE-1 CELLS (RIE-1 CELLS): These cells are from the laboratory of Dr. Ray Dubois (VAMC). They are typically cultured in Dulbecco's Modified Eagle's Medium supplemented with 10% fetal calf serum.

REOVIRUS: Laboratory strains of either serotype 1 or serotype 3 are used. They were originally obtained from the laboratories of Bernard N. Fields (deceased). These viruses have been described in detail.

RETROVIRUS: The U3 gene trap retrovirus used here were developed by Dr. Earl Ruley (VAMC) and the libraries were produced using a general protocol suggested by him.

SERUM: FETAL BOVINE SERUM Hyclone Laboratories cat. no. A-1115-L.

20

#### **Identification Tags for Isolated Nucleic Acids**

Genomic sequences, tagged with a vector, such as the U3 gene trap vector, are given a number corresponding to the genomic library of mutant cells from which the sequence was isolated., and a letter indicating a unique member of the library. More than one sequence with the same number and letter indicates multiple, unique sequences obtained from the genome surrounding the vector insert that "tagged" the gene. Such genomic sequences are obtained using vector-based primers, from which sequencing occurs 3' to 5' or 5' to 3'. In the former case, to recover the orientation of the gene into which the vector inserted, the sequence derived from the vector primer must be reversed and complemented. Such reverse complement sequences are designated "rE". In the case of genome sequencing from a primer that occurs 5' to 3' (i.e. the primer is at the 3' end of the vector), no changes are needed, since the derived sequence is the sequence as it appears



in the gene disrupted. Such sequences are designated "B4". Homologies indicated below each genomic sequence are in the positive direction, unless explicitly noted to be on the negative strand. As an example, SEQ ID NO. 27 comprises a nucleic acid sequence encoding a novel polypeptide on the positive strand, while the negative strand encodes ferritin.

	SEQ ID NO:	Lab Designation
	1	32-3-2#1E/-rE
	2	L191B2E#1-RE
10	3	L191B2E#3+-rE
	4	21-5-9E-RE
	homology to: emb/AL021154/HS15005 human DNA sequence	
	5	14A14E-rE
15	6	4cx-b4
	7	5a-b4
	8	6BSA12-B4
	9	X7B/B4
	10	x27b4f_1
20	11	12C#A-rE
	12	10-3b(5/2/96)/-rE
	13	10_4B_4-rE
	14	6BE60-rE
	homology to: alpha-trophomyosin	
25	15	19D3E-rE
	16	L19D16E-rE
	17	2b_rE
	18	14_24_#6-rE
30	19	7A7'-rE
	homology to: annexin II/dynein I	
	20	L12cx#6-rE
	homology to: gb:X51760 human zinc finger protein ZFP-36	
35	21	L12cx#11-rE

- 22 19D5E-rE  
homology to: 6-pyruvoyl-tetrahydropterin synthase (gb/M77850/RAT6PTHS)
- 23 12\_3b#7-rE
- 5 24 12\_3B#8-RE  
homology to: gb/AA871174/vq32a08.r1 Barskad bowel MPLRBg Mus musculus cDNA clone 10959265'
- 25 9B27-2-E  
homology to: RAT LOCUS RNU53922 04-MAY-1996; Rattus norvegicus DnaJ-like protein (RDJ1) mRNA, complete Cds, ACCESSION U53922 (on negative strand)
- 10 26 x15-rE
- 27 X11-rE  
homology to: ferritin H (on the negative strand)
- 15 28 X20-rE  
homology to: LOCUS RATGL5A Rat NICER element (GL5-14)5' long terminal repeat, Acc.No. M59028 M33535N1D
- 20 29 X4-rE
- 30 14A7E-rE  
homology to: MMSMAD7 3681 bp mRNA ROD 31-JUL-1998 DEFINITION Mus musculus mRNA for Mad-related protein Smad7 ,149 bases
- 25 31 14A13E-rE
- 32 14\_7#2E-rE  
homology to: N-acetylglucosaminyltransferase I
- 33 12CX#6-rE
- 30 homology to: gb|AA522204|AA522204 vf98g09\_r1 Soares mouse mammary gland NbMMG Mus musculus cDNA clone 851872; also 5' similar to gb X51760 zinc finger protein ZFP-36 (HUMAN), gb L20450 Mus musculus DNA-binding protein mRNA, complete cds (MOUSE); Length = 442, 925 bases (shares homology with SEQ ID NO:20)
- 35 34 12C\_2B#9E-rE
- 35 12CX#11E-rE
- 36 x5-rE
- 37 8C5\_11-rE
- 38 191E2E-rE
- 40 39 19\_7AE-rE

- 40 19\_9BE-rE  
homology to: LOCUS HS347M6 56583 bp DNA PRI 14-JAN-1998 Human DNA  
sequence from PAC 347M6 on chromosome Xq22, CSTF2 (Cleavage Stimulation Factor,  
CF-1, Polyadenylation Factor) 64 kD subunit gene
- 5 41 191E9E-rE  
42 191E8E-rE  
43 14C\_2E/-rE  
homology to: gb/H31084/EST104778 Rattus sp. cDNA - 5' end similar to signal  
10 recognition particle subunit(19kDa) (on negative strand)
- 44 14H1E-rE  
45 14G3E-rE  
46 14G\_2E-rE  
15 47 6\_3\_6\_2E/-rE  
homology to: Rattus norvegicus cis-golgi gp130 (on negative strand); and  
a HUMAN EST (on positive strand) A1127398; qb70g11.x1 Soares fetal heart NbHH19W  
Homo sapiens cDNA clone (1705508 3' mRNA sequence)
- 20 48 14H4E/-rE  
49 18A\_8\_4E-rE  
50 18A\_8\_1E-rE  
51 SCB2\_19E-rE  
52 L197B3E-rE  
25 53 L195C5E-rE  
homology to: H. pylori and C.jejuni
- 54 21\_5\_7E-rE  
homology to: id3 gene; emb|AL021154|HS150O5 Human DNA sequence from clone  
30 150O5; HTGS phase 1 [Homo sapiens]; containing the E2F2 gene for transcription factor  
E2F-2 and the ID3 gene for Inhibitor of DNA binding 3 (dominant negative  
helix-loop-helix protein), 1R2, Length = 133667, 971 bases
- 55 L195B1E-rE  
35 homology to: vK72b07.s1 Knowles Solter mouse 2 cell Mus musculus cDNA clone  
960181 5'
- 56 L194c4E-rE  
57 L193A1E#A-rE  
40 58 L192A3E-rE  
59 L1739E-rE

- 60 L192B3E#13-rE  
contains sequence identical to: insulin growth factorII/mannose-6-phosphate receptor
- 61 3 2 4 rE  
5 located in the same region of the genome as calcyclin, but the gene is "read" in the  
opposite direction
- 62 36 7 1 a-rE  
63 36 5 1 4 a-rE  
64 34 25 5a-rE  
10 rat satellite DNA (RATRSSID 93 bp, ROD 12-MAR-1984)
- 65 34 24-126/rE  
homology to:  
15 **HSU49928** (3096 bp mRNA) PRI 06-APR-1998, Homo sapiens TAK1 binding protein  
(TAB1) mRNA, complete cds, ACCESSION U49928 NID g1401125, and  
**HS333H23** (142274 bp DNA) HTG 17-JUL-1998 Human DNA sequence
- 66 34 23-1/rE  
67 36 5 2-6/rE  
20 68 36 5 2-196/rE  
69 34 23-3/rE  
homology to: gb|L16546|RATAP1X Rat P-glycoprotein (mdr1b) gene  
70 34 25 23-rE  
71 36 5 2-196/rE  
25 72 31 3 9/rE  
homology to: AA798638 568 bp mRNA EST 10-FEB-1998, vw34b06.r1 Soares mouse  
mammary gland NbMMG Mus musculus cDNA clone1245683 5, mRNA sequence, 824  
bases.  
73 31 3 6-2-rE  
30 74 31 3 17-rE  
75 31 3 5-rE  
homology to: AF046001 2347 bp mRNA PRI 19-FEB-1998, Homo sapiens zinc finger  
transcription factor (ZNF207) mRNA, complete Cds, 833 bases.
- 35 76 31 3 15#1/rE  
77 24 3 5#1/rE  
78 31 4 4#1/rE  
79 31 3 19#2/rE  
80 31 4 5#1/rE  
40 81 24 9 3#2/rE  
82 L24\_26\_1-BL

homology to: AI045472 396 bp mRNA EST 06-JUL-1998, UI-R-C1-jz-h-09-0-UI.s2  
UI-R-C1 Rattus norvegicus cDNA clone UI-R-C1-jz-h-09-0-UI 3', mRNA sequence.

	83	L24_26_1-B4
5	84	L22_5A1/rE
	85	L24_3_2B/rE
	86	L24 4-2/rE
	87	L24 5-2/rE
	88	L24 5-3/rE
10	89	(15-)L28AP/rE
	90	L24 26-10/rE
	homology to: LOCUS R06687 403 bp mRNA EST 03-APR-1995; yf10a10.r1 Soares fetal liver spleen 1NFLS Homo sapiens cDNA clone 126426 5'	
15	91	L24 26-2A/rE
	92	L24 26-2B/rE
	homology to: gb AA590026 AA590026 vm22g03.r1 Knowles Solter mouse blastocyst B1 Mus musculus cDNA clone 990964 , 459 bases, 139A; and Rattus norvegicus Eker rat-associated intracisternal-A particle element	
20	93	14 7#2E-rE
	homology to: N-acetylglucosaminyltransferase I; this sequence shares homology with SEQ ID NO:32.	
25	94	x18
	95	31_3_9-rE
	96	31_3_6_2-rE
	97	31_3_17-rE
	98	31_3_15#1-rE
30	99	24_3_5#1-rE
	100	31_4_4#1-rE
	101	31_3_19#2-rE
	102	31_4_5#1-rE
	103	24_9_3#2-rE
35	104	14XD#12E-rE
	105	70A-rE
	106	31-3-4-rE
	107	3_6_9-NeoG-rE



- 108 31\_4\_2-rE  
109 3\_2\_13-rE  
homology to: calcyclin
- 5 110 3\_2\_4-E  
homology to: pistlire-alpha 1 (with homology to calcyclin on negative strand)
- 111 L25-10/-rE  
homology to: calcyclin
- 10 112 L24-4-3/-rE  
113 L24-9-1-rE  
rat id sequence
- 15 114 17-L25-27#7-rE  
homology to: calcyclin
- 115 L21C1E-rE  
homology to: calcyclin
- 20 116 L24-5-3BE-rE.  
homology to:  
LOCUS H32572 310 bp mRNA EST 08-SEP-1995 EST107805 Rat PC-12 cells, untreated  
Rattus sp cDNA 5' end, ACCESSION H32572, and
- 25 LOCUS AA858747 470 bp mRNA EST 10-MAR-1998 UI-R-A0-bb-e-01-0-UI.s1 UI-R-A0  
Rattus norvegicus cDNA clone UI-R-A0-bb-e-01-0-UI, 3' similar to gb|AA473081|AA473081  
vd44b07-r1 Barstead MPLRB1 Mus musculus cDNA clone 803413 5' mRNA sequence
- 117 L24-4-2BE-rE  
30 homology to: LOCUS MMU51002 6495 bp DNA ROD 16-JAN-1997 Mus musculus Dlx-2  
gene, complete cds, ACCESSION U51002 NID g1477589
- 118 17-3-3B-B4  
119 L24-26-3/-rE  
35 homology to: RNU23776, DNA ROD 10-AUG-1995, Rattus norvegicus Eker rat-associated  
intracisternal-A particle element
- 120 12\_2B#2-rE  
121 05-17-3-3He-1-T7
- 40 122 21\_5\_8E-rE  
homology to: emb|AL021154|HS150O5 Human DNA sequence from clone 150O5;  
1p36\_13-36\_22, contains the E2F2 gene for transcription factor E2F-2 and the ID3 gene for  
Inhibitor of DNA binding 3(dominant negative helix-loop-helix protein, 1R2, Length =  
133667, 971 bases
- 45 123 X18H-t7

124 18A\_8\_4E-rE  
 125 L24-5-2BE-rE  
 126 L24-4-2AE-rE  
 127 L24-10-1BE-rE

5

### **Genes Necessary for Viral Infection**

Some of the isolated sequences disclosed here comprise sequence encoding the following proteins: tristetraprolin (human ZFP-36), 6-pyruvoyltetrahydropterin synthase, a eukaryotic DnaJ-like protein, ID3 (inhibitor of DNA binding 3), N-acetylglucosaminyltransferase I (mGAT-1), cleavage stimulation factor (CSTF2), TAK1 binding protein, human zinc transcription factor ZPF207, Dlx2, Smad7 (Mad-related protein), and P-glycoprotein (mdr1b).

### **Isolation of cellular genes that suppress a malignant phenotype**

15 We have utilized a gene-trap method of selecting cell lines that have a transformed phenotype (are potentially tumor cells) from a population of cells (RIE-1 parentals) that are not transformed. The parental cell line, RIE-1 cells, does not have the capacity to grow in soft agar or to produce tumors in mice. Following gene-trapping, cells were screened for their capacity to grow in soft agar. These cells were cloned and genomic sequences were obtained  
 20 5' or 3' of the retrovirus vector, i.e. SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, SEQ ID NO:36 or SEQ ID NO:94; sequences designated with an "x" in the clone name). All of the cell lines behave as if they are tumor cell lines, as they also induce tumors in mice.

Of the cell lines, two are associated with the enhanced expression of the prostaglandin synthetase gene II or COX 2. It has been shown that disruption of gene function by retroviral targeting of an upstream gene has lead to increased expression of a downstream gene product, COX 2. When a small molecule inhibitor of COX 2 enzyme was added, reversion of the transformed phenotype occurred. The COX 2 gene has been found to be increased in pre-malignant adenomas in humans and overexpressed in human colon cancer. Inhibitors of COX  
 25 2 expression also arrests the growth of the tumor. One of the cell lines, x18 (SEQ ID NO:94), has disrupted a gene that is now represented in the EST (dbest) database, but the gene is not known (not present in GenBank).

30

Each of the genes from which the provided nucleotide sequences is isolated (and all clones designated with an "x") represents a tumor suppressor gene. The mechanism by which the disrupted genes may suppress a transformed phenotype is at present unknown. However, each one represents a tumor suppressor gene that is potentially unique, as none of the genomic sequences correspond to a known gene. The capacity to select quickly tumor suppressor genes may provide unique targets in the process of treating or preventing (potential for diagnostic testing) cancer.

#### **Isolation of entire genomic genes**

10 An isolated nucleic acid of this invention (whose sequence is set forth in any of SEQ ID NO:1 through SEQ ID NO:127), or a smaller fragment thereof, is labeled by a detectable label and utilized as a probe to screen a rat genomic library (lambda phage or yeast artificial chromosome vector library) under high stringency conditions, *i.e.*, high salt and high temperatures to create hybridization and wash temperature 5-20°C. Clones are isolated and  
15 sequenced by standard Sanger dideoxynucleotide sequencing methods. Once the entire sequence of the new clone is determined, it is aligned with the probe sequence and its orientation relative to the probe sequence determined. A second and third probe is designed using sequences from either end of the combined genomic sequence, respectively. These probes are used to screen the library, isolate new clones, which are sequenced. These  
20 sequences are aligned with the previously obtained sequences and new probes designed corresponding to sequences at either end and the entire process repeated until the entire gene is isolated and mapped. When one end of the sequence cannot isolate any new clone, a new library can be screened. The complete sequence includes regulatory regions at the 5' end and a polyadenylation signal at the 3' end.

25

#### **Isolation of cDNAs**

An isolated nucleic acid (whose sequence is set forth in any of SEQ ID NO:1 through SEQ ID NO:127), or a smaller fragment thereof, or additional fragments obtained from the genomic library, that contain open reading frames, is labeled by a detectable label and utilized  
30 as a probe to screen a portions of the present fragments, to screen a cDNA library. A rat cDNA library obtains rat cDNA; a human cDNA library obtains a human cDNA. Repeated screens can be utilized as described above to obtain the complete coding sequence of the gene

from several clones if necessary. The isolates can then be sequenced to determine the nucleotide sequence by standard means such as dideoxynucleotide sequencing methods.

#### **Serum survival factor isolation and characterization**

5       The lack of tolerance to serum starvation is due to the acquired dependence of the persistently infected cells for a serum factor (survival factor) that is present in serum. The serum survival factor for persistently infected cells has a molecular weight between 50 and 100 kD and resists inactivation in low pH (pH2) and chloroform extraction. It is inactivated by boiling for 5 minutes [once fractionated from whole serum (50 to 100 kD fraction)], and  
10   in low ionic strength solution [10 to 50 mM].

      The factor was isolated from serum by size fraction using centriprep molecular cut-off filters with excluding sizes of 30 and 100 kd (Millipore and Amnicon), and dialysis tubing with a molecular exclusion of 50 kd. Polyacrylamide gel electrophoresis and silver staining was used to determine that all of the resulting material was between 50 and 100 kd,  
15   confirming the validity of the initial isolation. Further purification was performed on using ion exchange chromatography, and heparin sulfate adsorption columns, followed by HPLC. Activity was determined following adjusting the pH of the serum fraction (30 to 100 kd fraction) to different pH conditions using HCl and readjusting the pH to pH 7.4 prior to assessment of biologic activity. Low ionic strength sensitivity was determined by dialyzing  
20   the fraction containing activity into low ionic strength solution for various lengths of time and readjusting ionic strength to physiologic conditions prior to determining biologic activity by dialyzing the fraction against the media. The biologic activity was maintained in the aqueous solution following chloroform extraction, indicating the factor is not a lipid. The biologic activity was lost after the 30 to 100 kd fraction was placed in a 100°C water bath for 5  
25   minutes.

#### **Isolated nucleic acids**

      Tagged genomic DNAs isolated were sequenced by standard methods using Sanger dideoxynucleotide sequencing. The sequences were run through computer databanks in a  
30   homology search. These genes can be therapy targets particularly because disruption of one or both alleles results in a viable cell.

What is claimed:

1. An isolated nucleic acid comprising a nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:112, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, and SEQ ID NO:127.
2. A nucleic acid comprising at least 20 consecutive nucleotides of a nucleotide sequence of claim 1.
3. A nucleic acid comprising at least 30 consecutive nucleotides of a nucleotide sequence of claim 1.
4. A nucleic acid comprising at least 40 consecutive nucleotides of a nucleotide sequence of claim 1.
5. An isolated nucleic acid encoding the protein encoded by the gene comprising the nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5,



SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:112, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, and SEQ ID NO:127, or a homolog thereof.

6. A host cell containing the nucleic acid of claim 1 or 5.
7. A nucleic acid comprising a nucleic acid that selectively hybridizes under stringent conditions with the nucleic acid of claim 1 or 5.
8. A nucleic acid having a region within an exon wherein the region has at least 50 % homology with a nucleic acid of claim 1 or 5.
9. A nucleic acid having a region within an exon wherein the region has at least 60 % homology with a nucleic acid of claim 1 or 5.
10. A nucleic acid having a region within an exon wherein the region has at least 70 % homology with a nucleic acid of claim 1 or 5.

11. A nucleic acid having a region within an exon wherein the region has at least 80 % homology with a nucleic acid of claim 1 or 5.
12. A nucleic acid having a region within an exon wherein the region has at least 90 % homology with a nucleic acid of claim 1 or 5.
13. A nucleic acid having a region within an exon wherein the region has at least 95 % homology with a nucleic acid of claim 1 or 5.
14. A polypeptide comprising the amino acid sequence encoded by the nucleic acid of claims 1 or 5.
15. A nucleic acid comprising a regulatory region of a gene comprising the nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:112, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, and SEQ ID NO:127, or a homolog thereof.

16. A construct comprising a regulatory region of claim 15, wherein the regulatory region is functionally linked to a reporter gene.

17. A method of reducing or inhibiting a viral infection in a subject, comprising administering to the subject an amount of a composition that inhibits expression or functioning of a gene product encoded by a gene comprising the nucleic acid set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:47, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:60, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:72, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:75, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:90, SEQ ID NO:91, SEQ ID NO:92, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, or SEQ ID NO:127 or a homolog thereof, thereby treating the viral infection.

18. The method of claim 17, wherein the gene is selected from a nucleic acid encoding a gene product from the group consisting or, or the gene product is selected from the group

consisting of: tristetraprolin (human ZFP-36), 6-pyruvoyltetrahydropterin synthase, a eukaryotic DnaJ-like protein, ID3 (inhibitor of DNA binding 3), N-acetylglucosaminyltransferase I (mGAT-1), cleavage stimulation factor (CSTF2), TAK1 binding protein, human zinc transcription factor ZPF207, Dlx2, Smad7 (Mad-related protein), and P-glycoprotein (mdr1b).

19. The method of claim 17, wherein the subject is a human.

20. A method of reducing or inhibiting a viral infection in a subject comprising mutating *ex vivo* in a selected cell an endogenous gene comprising the nucleic acid set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:47, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:60, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:72, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:75, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:90, SEQ ID NO:91, SEQ ID NO:92, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, or SEQ ID NO:127 or a homolog thereof, to a mutated gene

incapable of producing a functional gene product of the gene or to a mutated gene producing a reduced amount of a functional gene product of the gene, and placing the cell in the subject, thereby reducing viral infection of cells in the subject.

21. The method of claim 20, wherein the cell is a hematopoietic cell.

22. The method of claim 20, wherein the subject is a human.

23. The method of claim 20, wherein the cell is from the subject.

24. A method of screening a compound for effectiveness in treating or preventing a viral infection, comprising administering the compound to a cell containing a cellular gene comprising the nucleic acid set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:47, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:60, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:72, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:75, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:90, SEQ ID NO:91, SEQ ID NO:92, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:105, SEQ ID NO:106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117,



SEQ ID NO:118, SEQ ID NO:119, SEQ ID NO:120, SEQ ID NO:121, SEQ ID NO:122, SEQ ID NO:123, SEQ ID NO:124, SEQ ID NO:125, SEQ ID NO:126, or SEQ ID NO:127, or a homolog thereof, and functionally encoding a gene product necessary for reproduction of the virus in the cell but not necessary for survival of the cell and detecting the level and/or activity of the gene product produced, a decrease or elimination of the gene product and/or gene product activity indicating a compound effective for treating or preventing the viral infection.

25. A method of screening a compound for reducing or inhibiting a viral infection, comprising administering the compound to a cell containing the construct of claim 16 and detecting the level of the reporter gene product produced, a decrease or elimination of the reporter gene product indicating a compound for reducing or inhibiting the viral infection.

26. A method of screening a compound for effectiveness in treating or preventing a viral infection comprising contacting the compound with the gene product of a cellular gene comprising nucleic acid set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:47, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:60, SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:63, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:66, SEQ ID NO:67, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:72, SEQ ID NO:73, SEQ ID NO:74, SEQ ID NO:75, SEQ ID NO:76, SEQ ID NO:77, SEQ ID NO:78, SEQ ID NO:79, SEQ ID NO:80, SEQ ID NO:81, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:85, SEQ ID NO:86, SEQ ID NO:87, SEQ ID NO:88, SEQ ID NO:89, SEQ ID NO:90, SEQ ID NO:91, SEQ ID NO:92, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:98, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID



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27. A method of suppressing a malignant phenotype in a cell in a subject, comprising administering to the subject an amount of a composition that inhibits expression or functioning of a gene product encoded by a gene comprising the nucleic acid set forth in SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, SEQ ID NO:36 or SEQ ID NO:94, or a homolog thereof, thereby suppressing a malignant phenotype.

28. A method of screening a compound for effectiveness in treating a viral infection, comprising administering the compound to a cell containing a cellular gene functionally encoding a gene product whose overexpression inhibits reproduction of the virus but does not prevent survival of the cell and detecting the level of the gene product produced, an increase in the gene product indicating a compound effective for treating the viral infection.

29. A method of screening for a compound that can suppress a malignant phenotype in a cell comprising administering the compound to a cell containing a nucleic acid functionally encoding a gene product encoded by a gene comprising the nucleic acid set forth in SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, SEQ ID NO:36 or SEQ ID NO:94, or a homolog thereof, and detecting the level of the gene product produced, an increase in the gene product indicating a compound effective for suppressing the malignant phenotype.

**ATTORNEY DOCKET NO. 01123.0004**

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## DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

(X) Original    ☐ Supplemental    ☐ Substitute    ☐ PCT

**As a below named inventor, I hereby declare that:**

My residence, post office address and citizenship are as stated below next to my name.

I believe I am an original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled "**MAMMALIAN GENES INVOLVED IN VIRAL INFECTION AND TUMOR SUPPRESSION**", which is described and claimed in the specification

[illegible]

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information known by me to be material to the patentability of the claims of this application in accordance with Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a) - (d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate relating to this subject matter having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATIONS: (ENTER BELOW IF APPLICABLE)			PRIORITY CLAIMED (MARK APPROPRIATE BOX BELOW)	
APP. NUMBER	COUNTRY	DAY/MONTH/YEAR FILED	YES	NO
PCT/US98/21276	International	08/10/98	X	

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States

provisional application(s) listed below.

APPLICATION NUMBER	FILING DATE
60/062,021	October 10, 1997

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose all information known by me to be material to the patentability of the claims of this application as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS (MARK APPROPRIATE COLUMN BELOW)		
		PATENTED	PENDING	ABANDONED

I hereby appoint the following attorneys and/or agent(s) to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

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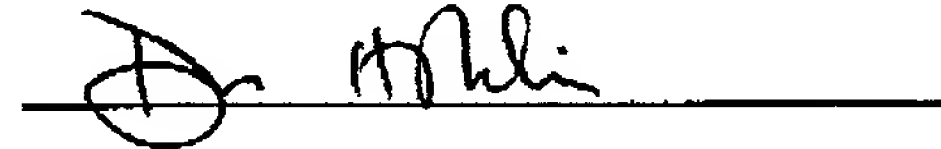
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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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-1-

## SEQUENCE LISTING

<110> Rubin, Donald H.  
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<120> Mammalian Genes Involved in Viral  
Infection and Tumor Suppression

<130> 22000.0086/P

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<151> 1997-10-10

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<210> 2

<211> 554

<212> DNA

<213> Rattus norvegicus

<400> 2

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gccaaattct	ccctcctatt	aaacttttca	cttcctttca	cttagtttcc	tnacttcttt	420
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<210> 3

<211> 891

<212> DNA

<213> Rattus norvegicus



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&lt;400&gt; 3

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&lt;210&gt; 4

&lt;211&gt; 974

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 4

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acgnnnctcc	caga					974

&lt;210&gt; 5

&lt;211&gt; 850

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 5

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tgnggagntt	gngaccttn	ttaccgnaan	gagtgggaaa	cgttttcctc	cgggttnang	120
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ggantaacag	ngnttgcntt	gtntngcnaa	acgaagagtn	tcctgnttgg	aataggngtt	360
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&lt;210&gt; 6

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<211> 531  
 <212> DNA  
 <213> Rattus norvegicus

<400> 6  
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 ananacatca gagatctctn gnacagtgtt tcacaagagt ctatcncana gagcacatct 180  
 gcccggggng anacacaact ctaaagtgtg ctcanntgat ctctctnttg tgtctctnac 240  
 atatgnggac atgtctctcag agtatnggnt ctcttgngcn cttntgcaca cacacacaca 300  
 cacacacaca cacacacaca cacncttctc tctggcacag ggntatggca nagcacatnt 360  
 tnnagantca nagctntata tgagtgtgtg gcgaaaggng tnatnanann gacnncccca 420  
 gcnnatatag gggggngnnc tctngggctc tcttnggnaa tntgngggng agtctgcnc 480  
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<210> 7  
 <211> 572  
 <212> DNA  
 <213> Rattus norvegicus

<400> 7  
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 aaccncgggg nctcntgttt tatttttaaaa aaaaagagtc ncatgtntat ttctctnatg 180  
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 tcacaacaca attcncgaga nattntgttc cncantggnn gtctnagntc ncatgttgtg 420  
 gngacangtt agnnccccc atnttncccc ccccttcaca ctgccccnag agagagaaan 480  
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<210> 8  
 <211> 906  
 <212> DNA  
 <213> Rattus norvegicus

<400> 8  
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 ganaganaga gagagagtgt gganctntnt cctcnganc ccanannan ngtgnggcnc 240  
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 acacagacac ntctctcang gggaacacat annagactnc acntgtgttt atattcnccc 420  
 ctcccnctca cacanacaca cacacagnag atattnngct actctctctc tgtcacaggg 480  
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 ctctcngaga angngagggc gntttacntt cccngtgccg tgnccnggcc cccgagactc 600  
 ccttngnac cccctntna accctctntt tgaaencaac ncacntccc cnttttctcg 660  
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 nccccg 906

<210> 9  
 <211> 914  
 <212> DNA  
 <213> Rattus norvegicus

<400> 9  
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 ttctcnantc tctaaaagng cnaaaagcgc ananacacgn gcctctctat anatctcacg 180

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ntctctgaga	tgtaganagn	gtcccctnct	cnatatatgt	gttnccact	ccnnaggng	360
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ccaccncnan	gacacaaana	naaggngtnn	anaaccccan	aaaaactnng	ntntcngntt	480
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ggngtggtg	ttngnntcaa	cacntntta	cctctctgnt	gnnanttgag	aaaatatattc	600
tntncttacc	cctctcccct	ctctgtgtgt	ngannatata	ngntctagat	gtcctnacc	660
tcccaaacc	tttctcnggn	agagacntct	ctntnttttt	ccccncttc	catttgaaan	720
anangagaag	gnccaaaaag	gngggngtct	tctcggaat	ncncctttt	ggccccccaa	780
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ccngaatttt	tttt					914

&lt;210&gt; 10

&lt;211&gt; 400

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 10

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gtgagatata	tgtgattctg	tggtgggtgt	ctcagagggg	gtttgggtta	ttggggataa	180
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aaaaaaaaaa	agggcccttc	ttgagtgatg	tccccttctt			400

&lt;210&gt; 11

&lt;211&gt; 880

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 11

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&lt;210&gt; 12

&lt;211&gt; 909

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 12

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ntnncgtang	tngantgacg	gaggccngna	atngggccc	gccanncnag	ggcgnagagg	660
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gaaaaagnc						909

&lt;210&gt; 13

&lt;211&gt; 927

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 13

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&lt;210&gt; 14

&lt;211&gt; 848

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 14

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tnatttnaan	gggngtagtt	tctggtnngt	tcattccttn	aaaaaaaaac	aaaacaaaac	180
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&lt;210&gt; 15

&lt;211&gt; 896

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 15

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ngngngnaaa	gagnnanttn	tttcaagggt	ccgnaacaaa	aagttgagng	angattccna	180
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aggatcnntt	tggacattg	gggtttgggt	agcntggnaa	cacgaaccct	tttgttcata	420
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&lt;210&gt; 16

&lt;211&gt; 858

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 16

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&lt;210&gt; 17

&lt;211&gt; 551

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 17

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&lt;210&gt; 18

&lt;211&gt; 888

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 18

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-7-

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&lt;210&gt; 19

&lt;211&gt; 867

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 19

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&lt;210&gt; 20

&lt;211&gt; 897

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 20

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&lt;210&gt; 21

&lt;211&gt; 435

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 21

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<210> 22  
 <211> 894  
 <212> DNA  
 <213> Rattus norvegicus

<400> 22						
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 <211> 594  
 <212> DNA  
 <213> Rattus norvegicus

<400> 23						
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<210> 24  
 <211> 586  
 <212> DNA  
 <213> Rattus norvegicus

<400> 24						
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<210> 25  
 <211> 909  
 <212> DNA  
 <213> Rattus norvegicus

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&lt;400&gt; 25

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&lt;210&gt; 26

&lt;211&gt; 576

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 26

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&lt;210&gt; 27

&lt;211&gt; 853

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 27

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&lt;210&gt; 28

&lt;211&gt; 825

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 28

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&lt;210&gt; 29

&lt;211&gt; 861

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 29

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&lt;210&gt; 30

&lt;211&gt; 149

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 30

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gaggcttttc	ctcccttcag	taggtcagc				149

&lt;210&gt; 31

&lt;211&gt; 857

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 31

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ttccaggngn	acangaacgg	gtgcggnggg	antaggggga	aangtttgga	gtgngccaaa	180
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aaggatttca	ggagatcctt	tgatttttat	tcgggtanga	cgttcangtn	gnggggattg	360
ggagcggana	accatttnna	cacaggattn	tatgaactat	ggtcanttgc	tttggtgtcc	420
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gnatntttgg	atcgcgtttc	gtgggatctt	aaaaaccggt	ttctgtgatt	ggcacgcaag	660
aaanactcat	gagctggtcc	ctggtgtgtc	tctcaggacc	aatcaaanac	ccatttccaa	720

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cggttttata	atgtctgggt	ctgtttgcac	aggaagcgaa	gtcacggctt	gcacccgtga	780
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caatcttcca	gttggtt					857

<210> 32  
 <211> 1630  
 <212> DNA  
 <213> Rattus norvegicus

<400> 32

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gnnttaannc	ttttnngngaa	tgttnacccc	aatnttcccc	tnaattttga	gtngngataat	180
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aaatcccgtt	gtttagatct	tgaccgtaat	caggagtcag	cgtagaggag	gccccggaag	360
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aggggagcgc	cgcgccgcn	tcccggggaa	ggacacattg	cttggttagca	ggaagccagc	480
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gcagggtag	ttgagacgat	cgggtgagct	tgggccgggg	acgccagcgt	cttcagtcct	600
ggggattgtc	ccaggagggc	aaggagcttg	gaggagggag	gccgcacagc	taggggagtc	660
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gaagagcctg						1630

<210> 33  
 <211> 883  
 <212> DNA  
 <213> Rattus norvegicus

<400> 33

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ggaatttggt	anaatttttn	taatggaaat	ngggcacttc	aattgggang	ataaaaacccc	180
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tgaatttgag	tgagtgcagg	tgaagtgaga	cttgggagna	caggtcatgc	ccaccaagg	300
gaggagcaag	ggntgggcag	tgtaggtggt	gnggtggtcc	ttcctggggg	gggcggggag	360
acagatgaga	acgttatttg	aggacaggca	caagtgttac	tgaaatgcaa	atccctgtag	420
atntggaaaa	gttctggntt	caggcttgat	gcttggggccg	gcaactgtgn	actttccctg	480
tacgttcagc	ccccccaccc	ttacgggaagt	tntcgtcact	gagantagtg	gctaatacaga	540
gtcttcaatg	gacctgcca	tcagaaagga	aggcgggctt	ttccgggtgc	ntaggtgtag	600
gattcgctca	gtagttaagc	agtcttaact	ggttntggct	gctgtgctct	ctgtcctgcc	660
gttggaattnt	ntgaggcatg	ttcaggcaag	ctccaaagtt	gcgacatggt	gagcacaggg	720
gcaggggggg	cgggcggacg	ggcaggggac	tgagcagtg	gagctggtgt	ggtgggtctt	780
tcccggggct	gagttggaat	ccgcggctac	ccgtgaggtc	ttagccactc	actagacca	840
gcggcagttt	ctgaataact	ttccttgtag	gggctgcaac	tct		883

<210> 34  
 <211> 913  
 <212> DNA



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&lt;213&gt; Rattus norvegicus

&lt;400&gt; 34

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ggttttntcc	naggggggga	gacccenttn	nccgcgggcc	tttcgnaatt	ttttggtcca	180
ccngtnaaag	attttcccat	ggcgcaccat	gtacgggttg	cgaggngtat	taggcggnaa	240
cggtttttna	gtgggcctaa	tacggnanat	aggaggacga	tttgtnttgg	tttgtngagc	300
cagtaccttn	gnaaagagtt	gtagttttga	tccggcaacc	aaccacngtt	gtagcngngt	360
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cttcgggttc	ngacgttatg	ttgtgtggat	tgtgagcgga	taacaatttc	acacagattc	480
cgatngtagt	ccaatttggt	aaagacagga	tatntttccc	ttcaaagaaa	acagaaaaat	540
acagaaacgt	taattttcaa	atctcnaatc	tttcntttct	tcttcnntca	ttcattcntt	600
cntttcttct	tctttctttc	tntctttctn	nagaggagge	atgctagggt	aacagtagct	660
catttttaaaa	tctggcacct	ggaattaatt	tagggacaaa	acacctttat	gcaaaaaaaa	720
gtttatgttt	ttccatggaa	cacagtaaaa	tcaaaattaa	aagaatataa	caaaggcttt	780
ggtgatttgg	taggattttt	tttttcctgg	aggggaaaac	agatgacttg	gaaagtgtta	840
ggaacatata	aagccccagg	gaaagaaaaa	cgtttgattg	gtattaatta	aaacactgct	900
aatatattct	aat					913

&lt;210&gt; 35

&lt;211&gt; 320

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 35

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ggttgggtaa	atgaagagga	gagacagagt	gggaagtcgg	cttagtggat	atggacttca	120
aatttgatga	acaagcaatt	caaataagta	tcgtgggctt	gactggtatg	aagaccggtt	180
tgcaaagcag	tgntcataag	agagaaaaga	gagagagaga	gagagagaga	gagagagaga	240
gagaaagaga	gagagtgtgt	gttggtgttg	ttgttgttgt	tgtttattgg	tttataacaa	300
gatntacntt	tggttaacttt					320

&lt;210&gt; 36

&lt;211&gt; 389

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 36

gggggggngc	naaaagggtc	tttcttttna	naaaaatcnn	gganggaggc	cncnanacgg	60
ctnttanann	tnntcngggt	gtncctcncc	gntgtgggga	atganatntc	gntctcgaca	120
tcaggggatt	ggagattntc	tgngctcncc	nctcacnacc	cagaagaagc	gcacagagan	180
cagagtanca	catcatacac	acctnttcag	ctacagagcg	antnctctan	aaggggactc	240
ggggganaac	acaaccctcc	tcctcttctg	actgngagng	cgcntgttag	gntctgtcta	300
cccancaagn	cttgtgcagn	ntgngaacct	ctctntgggg	tagagtgtgt	tgngggagca	360
gggcgtantg	ttccaggnet	agnctttca				389

&lt;210&gt; 37

&lt;211&gt; 882

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 37

agnaacgcgg	ncggnggnnc	tcnccngcg	gagcnggncc	nccccnngn	ncccagaana	60
gnagcgctcg	gngannnccc	acgngnagac	nnnggctgcc	ccncgngncc	anggcnttnn	120
ncnncccccc	cgnatccggn	ncnncccccc	ctccctnggg	gngcgggggt	ccngngccg	180
ngnggatacc	nggcganncn	ttgtgcccc	gcnngggggg	naggaccccc	ggcaccggcc	240
cngacccana	ncagnngctt	ngtggggggc	ccccccgcca	nagaacgaat	tnccgcnccg	300
gccgcggcca	tcggaacnnc	cctagcagng	cgtcntgcta	ggcnggnnna	cgggnatccg	360
caancccncc	cttngtaccg	ggacagccgn	gggnccgtat	gggctgngcg	ntngggccgta	420
gccanntncc	tttngaaang	acncggnagc	tnttcattccg	cctcacaac	cncngggncn	480
gngggggctn	tntcntgngc	cgcccgcgc	gtgngcgcan	aaaaaaaaa	aanncgccn	540
tcnccccctc	ttttggccng	ggtnccccgc	ncaccccggtg	cagagtnccn	nnccccccac	600
aacctcacac	cgatcccngt	gggttcccn	ngggagtcgc	ncngncnag	cnggnttctc	660



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cccatnncgc	gnngcttnag	cgngccnnnn	cacngtttgt	nngngnntgc	ctcccccttcn	720
tccttgaggc	aaaagcccg	acngtntctg	tggaccacnn	tgctgaggng	ctgggcgcen	780
cgntctctct	ctctctcnct	ctctctctct	ctctatctct	ctttctctct	ctggggcccc	840
tcccttgntg	nngccanaag	nnngcnnacc	cgtaaagtaa	gt		882

<210> 38  
 <211> 975  
 <212> DNA  
 <213> Rattus norvegicus

<400> 38

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agactaccta	taccgtcatt	aaaaactcct	caccagcatt	tactatgggt	ggactttcag	900
agatctcaat	caactctttc	ccaccagtc	tactgaaagn	ttccacctgt	agcggcccaa	960
gcaaactgag	atntt					975

<210> 39  
 <211> 850  
 <212> DNA  
 <213> Rattus norvegicus

<400> 39

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ggtttcaaac	aatgcttaag	ttgtggggag	aacnagnacg	tccgttceng	accngttta	180
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gagtgcagtg	caggcctcac	nttgccccag	ttantcagtg	cccacagagt	agcttttttg	720
agacgntagt	aaggtcttag	gggaagggaat	gtagtcgatc	cttctccttg	gtggccctca	780
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agtttcgagg						850

<210> 40  
 <211> 889  
 <212> DNA  
 <213> Rattus norvegicus

<400> 40

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nccgggaccg	naaccgtttt	tggccnaaan	ncgagaagtg	ccttccnggc	aaagtagggg	180
accaaaggtn	gggggagaga	attgggggtt	gtncagngtt	ccggttcnac	ggaaggagcc	240
ggttgttggg	attgtttcca	aggagnngt	ttngnaccgg	agcacctcng	ggnggaccat	300
ggggnttgcc	tggttagagac	cngcnggatg	ttttgggttc	gnattcgggg	agggatttcg	360
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gcagtttcgc	tgctgtcctt	tggcaatgng	cntgggnatt	ngtgggcaga	ngagattccc	480
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ggccacctgg	catcttgagg	tgggttgcca	atgagtgaat	gaatgagtga	gtgaatgaat	840
gaatgaatga	atgaatgaag	caagcttcag	ggagattttc	agagaagtg		889

&lt;210&gt; 41

&lt;211&gt; 929

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 41

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agttcattcc	tttgaccatc	ttttgnccct	ccatcttgta	aacanttaag	tctattgcat	300
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gaagcttggg	gcttatgttt	tgcagatcca	ttgtaattaa	aaaagaattg	taaggagatg	900
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&lt;210&gt; 42

&lt;211&gt; 943

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 42

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natcccaatt	aaggntnaac	nggtttaatt	tgtnntccnc	ntaccnaccn	ggtttnccgt	180
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ggaacccagc	nagtgcttgg	aggtaaaaaga	tcacttccnt	ntcccttagt	caggancntt	420
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tctctgaagt	caaggagccg	caccagcact	tcagttgtgg	gccataatca	agncangact	900
gaaagttgcc	acctgtagng	gccgcaagca	aactgagatn	ttg		943

&lt;210&gt; 43

&lt;211&gt; 867

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 43

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gttttgnngn	nattttcccc	ntgggtcggtt	tntgnacgat	ccacggttga	ccgacgacgn	180

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acggaccgac	aaccaanacg	taaaggggaa	ttgtggaggg	gttggaggtt	tagatgcccc	240
gacccaggac	gtgcggccan	cttccggaga	cccacctttc	ttgtnggccg	ggncggcgcg	300
cagcgnagcc	atttccaccg	gatccctata	gcngggccagc	ctagcaggcn	gaacaccagc	360
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ggacggtctt	cgcgcgagaa	gagtaccctg	cctttcaggt	gcgggagtta	cntcagcctg	540
ctgcacaccc	ggctgtgcgc	antcttctgg	tgtggccggg	acggttcacc	cagaggagtc	600
tctgtagttc	ggagcaagat	gtcgggttaa	tctggcagga	aaatgccttc	tatgctcatn	660
tatatattcc	tgtttccctc	agcttgcttt	cgacttagta	aggtaacatt	tcagagcggt	720
gcacttagta	ctttttggca	ctgtgctgta	taaatataaa	tgttccacac	ttaacatctt	780
agatgttata	tctaaagata	tgcattctta	aacttcgaaa	gcgcataccc	taaaatttca	840
tatttttgca	tacattggtc	agctgtg				867

<210> 44  
 <211> 303  
 <212> DNA  
 <213> Rattus norvegicus

<400> 44						
ggaaatgatt	agtccaagaa	atatttgagc	agaagggagt	taggggttttc	aaattaggaa	60
agtggaatcc	acagagttcc	cttgacagag	aataataaaaa	ggactctggg	gtgtcagaat	120
ggtgggcatt	aacctgatct	tccacttgag	ggtaagggaa	atgattagtc	caagaaatat	180
ttgagcagaa	gggagttagg	gttttcaaat	taggaaagtg	gaatccacag	agttcccttg	240
acagagaata	taaaaaggac	tctgggggtg	cagaatggtg	ggcattaacc	tgatcttcca	300
ctt						303

<210> 45  
 <211> 840  
 <212> DNA  
 <213> Rattus norvegicus

<400> 45						
aaaccggnng	aanaaaaaan	gaaanngang	gcnnnaaaaa	agttnnngaca	gaaaaaactt	60
tnggaaaaaa	gganggggan	aaggcaggng	nccnactnaa	aanggncttt	tcnaagnng	120
anagagntgg	naatnagnaa	naggacattc	ttinnaacctc	cnanggnngn	nggaannaat	180
ngggattgag	cngccaccat	tagggangaa	gttngaattn	ngggggcccg	gngagttaaa	240
angattcccn	ggttttttta	aacagagaa	acctncagg	acagatnaac	ccgagattgg	300
ttccctngaa	aattnnngan	aaagataaan	gtaggagcat	tcaaagtag	anggtaaaa	360
taatgggaga	catagacacc	aaaaaaagcc	agttcagtg	gccccgaagg	ngcattaagg	420
gaggaccagg	aaacggcagc	anagccacng	gcagccgcct	gccccnacac	cagtnattcc	480
cgcacntaga	tccagcgcnt	ggggggcggg	cgggggcgcg	ntgngcagng	aagntnnngc	540
gcaacaantt	tgcntagacc	ggntggaacc	ggttagaacc	ggccgcgcgc	gaccggccgc	600
ccgttccgga	ttntgcgttc	acaaagggag	gcgggactca	cgacntngt	atcnttngg	660
tcccaacccc	ggcccccna	cccnacccc	nttgctccctg	tggcattegc	gttctttccg	720
ccgtctccct	cgcgggccgn	ttntctgcgc	ctggtgatcc	tttcgccatg	gtcctntgga	780
gaaagaaaaa	atctttaatt	tnctagggac	gtccttttcc	tgtagtcgta	attgtagaaa	840

<210> 46  
 <211> 893  
 <212> DNA  
 <213> Rattus norvegicus

<400> 46						
gagaaggann	aggnggggng	agngaagana	gaggagggaa	gaaangaagg	tggaganaag	60
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gnagnatagg	agaggaaaga	aagganggan	agaagagaaa	agaanganga	gagaaaggaa	180
agaggaaaga	aagaggggag	aagaggaaga	aanagaggag	gggangagag	ggaggataag	240
agaggaaaga	gggaganagg	nttgaaaagg	gaaagagaag	gagaaaggna	gnaggngngg	300
aagagaggna	aggagagagg	gganaanggt	aagggggnaa	agaangagaa	gtatnggggg	360
aaaggaggag	angaaagaag	aaagaganga	ggaggagagg	gagagtggag	aataaagggg	420
agggaaaagg	angagaaaga	gagagaggg	gagggagaaa	nagagaagga	tagnggggtg	480
gagaaggaga	aaggagagaa	ggagaaggng	agaggagaa	tgaagaagga	gggagtaaga	540
aaggantgag	naggaaagga	ganagagagg	tagagagaaa	anaaagagg	aaanggagg	600
gaggagggng	nanaaggaa	agaggngnga	aanangagag	aggggaaang	gggaaggaaa	660

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ggaggaaaaa	aagnagagaa	gaagagnaat	gggaaggang	nagtagnaaa	agaaaagnag	720
aggggagagg	gggangangg	ggganacggg	ggggaanaga	aaaagtgaag	gaggccccc	780
nacccccccc	ccccacacac	acacacagcc	ttttcgccgg	cggaagtgca	ggttggtcca	840
ggagcctgtg	gtcaatccag	tcagtagtgg	gcgaggtgta	acatctgtgt	ccg	893

&lt;210&gt; 47

&lt;211&gt; 789

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 47

taaaananng	gnngannanc	tnnaaaaaan	tntcttngga	attnncagga	nggaggntaa	60
tngggcgggc	ancatcaatg	gtanaaat	gggggggnng	annaaatca	tnaanncaac	120
cgtttccana	gncaaccatt	ctgggngncc	caaggttnga	ngagntccgn	tcaaggngaa	180
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gctgagaaga	ttcgttgggg	gacccaggag	tgaaggtttt	tnacctgtgt	ntntcgggaa	300
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agcgtttact	ttttnttgnc	cgcagccaat	ttgttntgct	tggtttcttc	ngaattcccg	420
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ttccggcagn	tacgantctt	caacaagagc	cagagaaggc	gggtgcagag	nttcattagg	540
acgntcggaa	acccggcgtg	acttactttt	tccaagccca	ttggttgatg	agaatgatga	600
ctgacagggg	ggcgtggtca	cgctgtcgcg	ggcgggagcg	acgggtggag	ttaacgacga	660
aagctgcgcg	cgcagccatg	acccttcaca	gccacntatc	ggagggaggg	gcgggacagc	720
tttagcttgg	tgcgtgcgca	gccggacgtg	aggcagttgg	tggctctcca	tcgtcgattt	780
ctggttacc						789

&lt;210&gt; 48

&lt;211&gt; 872

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 48

gggggnnggct	tttttnggag	gcatanatng	gggnnngtcc	ggnaaacccc	attgggtcggc	60
cggggaagga	aaanggggct	ctnaaaatan	gttantggga	tggngcctta	agggggggcc	120
catgngccag	gaangcagat	tcaaaaatgt	tccaagtggg	aaaccanggt	tggnanaggc	180
cctccnggnc	gtnaaggagg	agaggagaga	tggagtttca	ggtgtgtttc	ccaccagtg	240
ttcccaggga	acacaaaacg	gatagggtcac	cntcaatgna	caaggaatta	aaagcttggg	300
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caccagagg	ttcataggga	gggcantatt	aggggtgtgc	ccttgtgaga	ggaagtgtgg	420
caengtgggg	ctgggtttga	gatntcagat	gntcaagcca	ggccattnt	ntctctctca	480
gtntctctcg	gtctctttct	cngtctctnt	tcagtctntt	cagtctctct	cagactctct	540
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caccangacg	ataataggct	aaacttntga	actetaagcc	agcctcaatt	aaatttntan	720
gagtcaaacc	agcctcaatt	aatgtttttc	atttctatga	gtcacagtgg	tcatggcatt	780
tctttacagc	aatagaaacc	ctaactaaga	cttgccgaaa	cctcaaccac	aacttcagcc	840
ctcagaagcc	caagagggaa	aagaccttga	at			872

&lt;210&gt; 49

&lt;211&gt; 785

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 49

tcgtaanttt	tnatccaccn	gtanangatn	ttccatgcc	ccatgtacgg	ttacgaggng	60
tatagcgtgn	acngtttttg	agtnggctaa	aaggaaatgg	agacntattg	tnttggtttt	120
gtgaccata	acttcggaaa	ggttgtgttt	tatccggcaa	caaccacngt	gtagcggtgt	180
tttttggttg	cagcagcaga	taacgcgcag	aaaaaggatn	tcaggagatc	ctttgatatt	240
ttnttcgggt	tctgacgntc	atgttgtgtg	gaattgtgag	cggataacaa	tttcacacag	300
aattcaaagg	agaggagcca	atatagaggg	ggaaaaaaa	agaaggggaa	agcattagtt	360
taaaaagttg	agagaacaaa	gtatgttttg	cttggtatgg	caaccaaaga	agcntgccag	420
gaatggtcgg	taaaaaggtg	aagagtcatt	aaacgtcttc	tgtccaaccg	ttaccggaaa	480
catgcaagga	atttcttaga	ctggccagga	ttggattgtg	ggaaaggtct	cttcaagcnt	540
ccccttggtg	tttatggcaa	gaaaatagtg	cggactatag	agagcgctcg	tctcaaagct	600



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tgtccccaat	agcagaaaag	cattgtccta	aattccttaa	aaggcaccgt	gaaataaata	660
ttacgaggac	acgatggcac	aagaaggagc	tttcaactct	gccaccagaa	cagttatact	720
tcatagtaac	catgttgccc	tgttcaatga	caaggcacgc	tctccagcag	aaagggaaaa	780
ggagc						785

<210> 50  
 <211> 889  
 <212> DNA  
 <213> Rattus norvegicus

<400> 50

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ggctttgggn	gggntttcat	anggnntgn	tttacccaat	tcagtttttt	attggtnntt	120
natgggcgca	gggatagnn	gttcnggntt	cccacangaa	tttgattnt	ggaatcacia	180
gtnaccagtn	gccgnaatca	cgagtttgcc	gctttntttc	ctacctana	ttcataatan	240
gaatgagtan	ttttttttta	ttgagnaang	tttnnacagg	tttagtaaac	atgaggacag	300
aggttttaag	ttgangatta	ggaaggagag	ttccggggga	cagaatgtgt	gtattntcag	360
tcagtgcact	accggaaga	gttgcagtc	ggttgaggaa	gggagcggat	ttcctggagg	420
ttttaaccaa	cagagagaaa	aagcatttac	tactgattaa	gcacacaatc	tctggattca	480
gagaagggtg	tttaccttta	tataaaatgt	ctcctaactg	cgtgactgtg	tgactttgtt	540
gaagtcaact	gagcactgac	tgtgttgtgt	gcaacatgg	aagaggacca	actttnttct	600
taaattttat	ttattattta	tgtcacgtgn	acacttggtg	cttttggttt	tgttctaatt	660
ttatctgcat	atatgtctgc	ataccacgtg	catttctgat	gcntacagat	gccagaaaag	720
gacaccgagt	ttcccctggg	antggagtta	tagatgggta	taagtctctg	agtaggtact	780
gggaagtga	cttcagtttc	ctctggaagg	gcagaaagcg	cttttcaaat	gctgggccat	840
gtatttcagc	ccctacttaa	tttataattt	tatttttagag	gatgtgctc		889

<210> 51  
 <211> 947  
 <212> DNA  
 <213> Rattus norvegicus

<400> 51

anaaaaatng	agaagangag	accccagaga	agaagnanga	gaganaacag	agaagaagag	60
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aaggagaaga	nggaaggnta	acataggagn	caagaatana	aaganaaaaa	gaggtagaga	180
anncagagaa	cgagaaaaga	tgaanaaaag	antanaangg	aagaaagang	nccagnanaa	240
anaaggcaga	aanaagatgn	cgtaaaaana	gagagaagat	aggnaaaata	gaggagaagg	300
ccnaacagga	ngggaagagc	agcgaattnn	agataaaacc	ggagganagn	nagagaagg	360
agagntngnn	aaggcaaaga	cagnanngag	nacggtaent	gccccagaag	gnngaagaan	420
gncnagangg	tgagggngng	cacngncnt	tccccttagg	aggncgccc	cccagagatc	480
aggtttcnag	gncaccgagt	tggatacnag	attatncacc	naggcaggaa	angantatng	540
caaaangatt	cgggngggg	tcacgggggtg	agaaataaan	tcannaaana	gaggacgngg	600
aggagggngg	gaaactctng	acagaaatng	caagcangaa	gccagccnca	ccaagcccc	660
nacngaagca	gcngagangt	tgcanngcgg	naggtccaaa	tcancgnagt	catggagnga	720
gcttcggng	ggcccnaganc	cantgaggaa	gggcaggaaa	ccatatacnag	ccgagccnng	780
ngangntgc	cctganacac	ccggagaggt	aattttttatt	tnacgggaag	cgtccagnca	840
agttcgtggg	ccggaagaga	cggtacttta	gtatacanag	ctnntgctnc	gagttgtng	900
nccttntnat	gnnagatctc	acaaangaag	ctnanaagta	gatatgt		947

<210> 52  
 <211> 860  
 <212> DNA  
 <213> Rattus norvegicus

<400> 52

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tttaaaatta	aaggggnggg	angtttttcc	ggttctattn	ngccnatctg	gggttacact	120
tttatccanc	ntttgntttt	ttanccggcc	gggttaaaaa	tgggggggga	ttagttcggg	180
tagngttn	cnacagcaca	gccctgtttn	tcttcgttcc	ngaaaaaaa	aaattttgct	240
ggtntcacia	ttttnttaaa	caggatttnc	ttcaaccatg	gattaataca	tttccggtgc	300
agnttgccc	gtttgttttt	tggntggata	gggatgccag	caggattcag	ggatgcccat	360
tgtgnttagt	ntctggccct	ttaggagagc	tttgggctaa	ttatgtgacc	gattttaaga	420
agtgggtgtg	ttgtgggtcc	agggactcac	ggatcagcct	ttattttata	aggacactgt	480

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ggaggagaga	cagaggctga	gctgcattct	gatgtcattt	gtgctgctgt	ggaagttaaa	540
gaaaagctgc	agaagtcagc	aaaacagatg	aataccaaga	agggcagtgt	gagtacagga	600
atggagagaa	aagtcagagt	ccagcttttg	ttaactccct	aggatcagac	anttctgcgt	660
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ggcaggcagg	aaatggaaca	tttaactgga	aacactgaac	ccacccatgg	caaacttagc	780
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gagtttgagg	ctagactggt					860

<210> 53  
 <211> 191  
 <212> DNA  
 <213> Rattus norvegicus

<400> 53						
aggtctgacc	acttggaagc	ttgccctgan	tcatagatga	gccactgtct	tcttcccctc	60
aattcctcag	gatggggaac	agccattggg	cttttagtag	aggagggaca	ggcccttttg	120
cagcaacagt	tctcccctga	atgttggtgc	tccacctata	cacatggggg	acttagcctt	180
atggatgccc	c					191

<210> 54  
 <211> 988  
 <212> DNA  
 <213> Rattus norvegicus

<400> 54						
ttnttggnna	cgggtntccg	nantatgaan	centtcccgg	ggttttttaa	aancccnnga	60
tattcgggga	tttgggtttt	nnacggcctt	tttttnagag	gccaaatncc	cntntnaang	120
cctttttatcc	ttecntttnt	gccccncttc	naattaggaa	gcntggtttg	nccgantntt	180
aagggttttta	gtentccttc	gttnntnttt	cccttntttt	ttccctnaag	ttataaagcn	240
ggtatntggt	ttgccaggnt	tctnttgtag	ccgtcatngc	gggttnccgn	ttaccagagn	300
tttgttcctn	ggccggtncc	ttccaatttt	ggantntccn	ggtcngnggt	ccnattncct	360
tgnaacngtt	ccacacntna	tgacaattaa	ttgtttcctg	tgtaatttgt	ccccggactt	420
ntggattctt	gngancaggg	cctntgtttc	atggaagcaa	actcccttaa	ntatttacca	480
ggttgattga	ttaagaaagt	antcatgnnt	gggaaaccca	cntgttttnt	tcccaggatg	540
gaancccgag	attttggaac	tgcagaggct	tcaggggtctg	ggaagcggag	gcaggcaaaag	600
aatggagtgc	actgtccttt	tgcaatatgg	ggtttgcctg	cctgctggct	cctctcntgc	660
tntctcagat	ggtgactgag	gctacttcag	caggactagg	aataatcatg	tccaggtggc	720
tgcccttccg	agcagaaagg	gacagacgtg	gggcgatgaa	gttgctatcg	tttttttttt	780
tttctgcaca	gactgcaaag	tgtgcagagg	gagggagggt	gtgcaaaaaa	aaaaaaaaaa	840
aaaaaaaaaa	aaaaaaccga	ggacgcagaa	gtagactgc	tgacccattt	ggtgcatgtg	900
tgcccatgga	gggaggggac	cttctcaaaa	gggttcacgc	agcangcatt	gaaagtnccc	960
cacntgtagg	gncgcaagca	actgagat				988

<210> 55  
 <211> 665  
 <212> DNA  
 <213> Rattus norvegicus

<400> 55						
gaaaaagatt	caggaanctt	atttttntcg	gttcgacttc	agtnngggaa	tgggcggana	60
catttcacac	ggatttgtaa	anacngtnac	ngaaacttgg	nggttcgtag	atccactttt	120
tttagacctg	agagtagttt	ttaaaatatt	tnaattaaag	gtttcctgca	cccacttttt	180
tttttatccc	taactttttca	tccagtatgg	tttttcaata	tcacanttta	atctaggact	240
ccttgcttaa	agcaattaca	agttaaatta	aaagtaagag	atggctnata	gctctcatta	300
ctgggatgca	ggtgtgaaac	aagtgatttg	tgtagaagct	ggcaggatgg	gtataaacia	360
gaacacgtgc	ccagaggatg	tattgaaagt	tggatttaag	tctctgagta	gtttatgcta	420
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gaggaagtgt	gagggagagc	attagatggt	actgttgggg	atttggaag	gccaggaaac	540
gttactccat	gcccaggag	ggtaggagaa	aggtttgggc	ttagctttga	ggacggaggg	600
aactggtggg	tggatatgag	gatgggtatg	ctaaaagcag	agtggttttc	aactattggt	660
cttct						665

<210> 56  
 <211> 857



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&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 56

aaaaaaagaa	aggaaagggg	agananaaaa	annangngan	aaaanagana	ganagaggna	60
agaggaagng	aggnggaaaa	gagaggagan	aaanaagagg	aaggagaann	gaggaaaang	120
aaaggaacaa	aaganaagng	anggaagana	aaggggagaaa	aaanaagagg	gagaaangga	180
ggaggggaaan	agagaanaga	gggggagaga	anncagagaa	nagaannngag	aaaaggngga	240
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gaagangaga	agangagnag	aggaaganga	ggggaagaag	aagaggngga	ggagaagaag	360
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ggaggaggat	actanggagg	ttgtttcaat	aaaagagngg	gatntaagat	taananaagn	480
aataatgccg	gtttntatct	gttcgggggg	ggtccttggt	ctccaaacac	aganntgggc	540
cagtttntca	aaattnaant	gngaagattt	cttgngntga	gagcagntca	gattnantng	600
nattnttttc	tagtttttna	cacaancttt	gtgntaacia	agagnganga	ttcnaggana	660
actcgnnttt	ntttgggagg	agactttggt	cctttcnatg	aagatgcagg	acngngaaga	720
cgcagggtgt	gaacaggaca	cagnnacgct	tnngtntntg	tcngentcag	cngcgtggga	780
atgagtcaga	gcagcacggg	gaggtgcctg	gatntaagct	ttctggtagg	gagaacagag	840
tgcaggcngc	ggcccag					857

&lt;210&gt; 57

&lt;211&gt; 902

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 57

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gncgaattta	anaagggggt	agggaaaaa	aaaacanaat	attccntcct	tagccatnaa	120
ccgaacttcc	ngcaaggaaa	aaaaatttgg	ngggngtaaa	gggcaccncn	tcccacaaaa	180
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gggatttngg	ggatttnaaa	atcngngttt	nnggcagggn	atccngaagt	tngaatecga	300
cgncnaccct	ttatttnagc	agttatttan	gggaacatgg	gagggnacca	tttcaaacca	360
nggatcgggc	cnggagtntg	agtgttcage	ccacngcctt	cnaacantac	cgggataagt	420
ttttccctgn	gccagagacc	catccangtt	ccagcaaaa	gntgggtcat	tngggcnagc	480
tccnngagtc	atcnnngggt	tctcccagcc	ngggggccat	ggtcgaaggc	aggttntttt	540
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gtcatntcng	gctagctccn	ttagctccnt	gtccagggga	cttcctggca	ctggattagt	660
ggnggactca	ggcttgcttt	tttttcagga	gaggttagat	tactaatcat	tcagatgttc	720
ataagtcaga	acactgagca	aagcaatagn	ttctcctcca	cntactgant	cacacgtgca	780
caacagccac	acccgcaatg	cttntaggag	caggtccagn	gnacttttgt	tttaactatt	840
tntggctctt	tattaatcag	cacataaata	cgcttcggtt	ctcctttttc	aatatgnatg	900
tg						902

&lt;210&gt; 58

&lt;211&gt; 852

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 58

acagaggggg	ggggggngtg	gaatttttng	naggangttn	tnggaaggcc	nctaaaaaag	60
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atttttnnnn	tattnaggat	caggataaat	angaaaangg	gnanattttt	nnnangnggg	180
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<210> 59  
 <211> 884  
 <212> DNA  
 <213> Rattus norvegicus

<400> 59

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gggggtatcct	aggggggtana	nttccaaata	nngataanaa	ttttttttaa	aattttaattg	300
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<210> 60  
 <211> 955  
 <212> DNA  
 <213> Rattus norvegicus

<400> 60

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<210> 61  
 <211> 1107  
 <212> DNA  
 <213> Rattus norvegicus

<400> 61

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tgaggggggac	tgggaacaat	actgatgctg	ttgccctcta	gtggcaaggt	caactccaag	840
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<210> 62  
 <211> 92  
 <212> DNA  
 <213> Rattus norvegicus

<400> 62						
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<210> 63  
 <211> 209  
 <212> DNA  
 <213> Rattus norvegicus

<400> 63						
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agttcccagg	cgaganttct	ttgtacaggg	nnccctctga	anncnctga	aagatttcac	180
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<210> 64  
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 <212> DNA  
 <213> Rattus norvegicus

<400> 64						
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 <212> DNA  
 <213> Rattus norvegicus

<400> 65						
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<210> 66  
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 <212> DNA

## &lt;213&gt; Rattus norvegicus

## &lt;400&gt; 66

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## &lt;210&gt; 67

## &lt;211&gt; 815

## &lt;212&gt; DNA

## &lt;213&gt; Rattus norvegicus

## &lt;400&gt; 67

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## &lt;210&gt; 68

## &lt;211&gt; 1034

## &lt;212&gt; DNA

## &lt;213&gt; Rattus norvegicus

## &lt;400&gt; 68

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gaaagagatn	tttc					1034

<210> 69  
 <211> 186  
 <212> DNA  
 <213> Rattus norvegicus

<400> 69						
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tcctgagcaa	gatggaaatt	ttacttggtc	tgttaactag	cgtgcattga	atggangaca	180
tatgat						186

<210> 70  
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 <212> DNA  
 <213> Rattus norvegicus

<400> 70						
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ntatcttgaa	atttcccagt	ttttnaattn	tattntaang	atattntagt	tnnaatttna	240
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<210> 71  
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 <212> DNA  
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<400> 71						
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gggttaagtn	tttncccaaa	agttggtttt	tnnaaaaanc	ccctttnncc	cggacgtttn	180
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<210> 72  
 <211> 824  
 <212> DNA  
 <213> Rattus norvegicus

<400> 72  
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 cactataaag gaggccagg ccaaggactg gcctcctctt gctcacgagg tcagacgcga 780  
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<210> 73  
 <211> 774  
 <212> DNA  
 <213> Rattus norvegicus

<400> 73  
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 <213> Rattus norvegicus

<400> 74  
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<210> 75  
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 <212> DNA  
 <213> Rattus norvegicus

<400> 75  
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&lt;210&gt; 76

&lt;211&gt; 880

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 76

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&lt;210&gt; 77

&lt;211&gt; 864

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 77

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&lt;210&gt; 78

&lt;211&gt; 874

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 78

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&lt;210&gt; 79

&lt;211&gt; 886

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 79

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&lt;210&gt; 80

&lt;211&gt; 865

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 80

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&lt;210&gt; 81

&lt;211&gt; 859

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 81

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&lt;210&gt; 82

&lt;211&gt; 1021

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 82

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&lt;210&gt; 83

&lt;211&gt; 1013

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 83

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&lt;210&gt; 84



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<211> 1002  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 84

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<210> 85  
 <211> 1031  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 85

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<210> 86  
 <211> 1039  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 86

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&lt;210&gt; 87

&lt;211&gt; 1058

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 87

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&lt;211&gt; 1043

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 88

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&lt;211&gt; 454

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

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&lt;400&gt; 89

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&lt;211&gt; 873

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 90

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&lt;211&gt; 876

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 91

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&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 92

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-31-

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&lt;400&gt; 93

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<210> 96  
 <211> 774  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 96

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<210> 97  
 <211> 248  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 97

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<210> 98  
 <211> 880  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 98

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<210> 99  
 <211> 864  
 <212> DNA  
 <213> Rattus norvegicus

&lt;400&gt; 99

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caggaantga	tcctnttggg	ttacagtcac	tttagcatag	gntgacagtt	ggngaccaan	180
tnatcttgcc	gtgttggaag	gagaggggan	taaggntgaa	gctcttgagt	ccnttgange	240



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&lt;210&gt; 100

&lt;211&gt; 874

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 100

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&lt;210&gt; 101

&lt;211&gt; 886

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 101

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&lt;210&gt; 102

&lt;211&gt; 865

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 102

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&lt;210&gt; 103

&lt;211&gt; 859

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 103

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&lt;210&gt; 104

&lt;211&gt; 883

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 104

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&lt;210&gt; 105

&lt;211&gt; 987

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 105

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&lt;210&gt; 106

&lt;211&gt; 1031

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 106

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&lt;210&gt; 107

&lt;211&gt; 1138

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 107

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 <213> Rattus norvegicus

<400> 108

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 <213> Rattus norvegicus

<400> 109

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<400> 110

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&lt;210&gt; 111

&lt;211&gt; 1069

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 111

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&lt;210&gt; 112

&lt;211&gt; 1058

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 112

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<400> 113						
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 <212> DNA  
 <213> Rattus norvegicus

<400> 114						
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 <211> 913  
 <212> DNA  
 <213> Rattus norvegicus

<400> 115						
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acaggattnc	ttggccntnt	gaaacacgta	tttatgtcag	gnttntaccg	tngcaaantt	240
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&lt;210&gt; 116

&lt;211&gt; 1123

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 116

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&lt;210&gt; 117

&lt;211&gt; 1116

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 117

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1116

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 <213> Rattus norvegicus

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 gtgggagacg tcttgccctc ttgctgctcc tattggagaa gtgcttattt ctggttctgg 420  
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 <212> DNA  
 <213> Rattus norvegicus

<400> 120  
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 caaccccaaa ggggaaggga anggaangga aaatnaattt ccttttnaaa aaggagnaaa 180  
 tncgggtang gaaaattccg gtgnggggtt ttcaaagggt ccccccggn ggnntaaaaa 240  
 attgaagttn antcnngggg ggggaaccaa nagaatataa anaaaccggg gtttccccc 300  
 gggagttcct tgggggtttt ccggttcgac ccgncgntta ccggaacct ntncctttt 360  
 tcccttgggg nagggggggg 380

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 <211> 998  
 <212> DNA  
 <213> Rattus norvegicus

<400> 121  
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 ccgggggag gaggtacca cctccccacc ctctctctcc ctctctcag gagcttatct 180  
 atcgggtgagc agcaagtagg aaaaggtaag ctgagaaaga gcacttggct ggctacagga 240

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cctcagcctg	aggtgtgaaa	caggagactg	ggcactgggg	aaacagcagc	actggctggg	300
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cttgggctgg	gtgctggaat	cctaaccctt	cagtgatggg	ggaactctgc	tccagtgagc	420
tgaagtatac	atgtggggaa	ttgggggggtg	gggtaggggg	aaggcaatcc	aaaggtcact	480
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acaagctnnt	atgggcaagg	gctttgggtc	nagnaagg			998

&lt;210&gt; 122

&lt;211&gt; 970

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 122

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caactgagat						970

&lt;210&gt; 123

&lt;211&gt; 884

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 123

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acattcaaac	tgtagtaagt	gttttaattt	tctactaaac	aataaaacct	ttagattttc	480
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&lt;210&gt; 124

&lt;211&gt; 855

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus



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&lt;400&gt; 124

cccccttcgg	ggggtttana	anggaatnaa	tgggtntntn	ccaggggggg	aaacccttna	60
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&lt;210&gt; 125

&lt;211&gt; 1059

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 125

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aaccaacaa	tcaacggnta	ttgtttcagg	ganttnttgg	taccaggcnn	ttggttttga	180
naanacggta	ggtcggggaa	gcnttgacgg	taagcccnng	gganaagggc	caacggngat	240
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&lt;210&gt; 126

&lt;211&gt; 1042

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 126

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ccctttttnt	naatcnncat	ngagggttct	aantgaacct	naaggnnatt	taacttttna	180
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caagtattga catggactat tnttaaggcc aaangagagg tcnccagaaa gttatacatg 1020  
taggttggcg cgctgaaagg at 1042

&lt;210&gt; 127

&lt;211&gt; 960

&lt;212&gt; DNA

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 127

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caagggagaa	acaggtcaga	ggtagagaga	gctcagtcca	gggactcacg	gtgaggtccc	900
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